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(54) Title: A METHOD OF NUCLEIC ACID SEQUENCING

(57) Abstract

The present invention relates, in general, to a process of enzymatically synthesizing nucleic acids containing nucleotides that are resistant to degradation. The invention further relates to methods of utilizing such nucleic acids in DNA and RNA amplification and sequencing, gene therapy and molecular detection protocols.

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A METHOD OF NUCLEIC ACID SEQUENCING

This is a continuation-in-part of Application No. 08/300,265, filed September 2, 1994, which is a continuation-in-part of Application No. 08/115,690, filed September 3, 1993, now abandoned, the entire contents of which applications are incorporated herein by reference.

TECHNICAL FIELD

The present invention relates, in general, to a process of enzymatically synthesizing nucleic acids containing nucleotides that are resistant to degradation. The invention further relates to methods of utilizing such nucleic acids in DNA and RNA amplification and sequencing, gene therapy and molecular detection protocols.

BACKGROUND

Cycle sequencing polymerase chain reaction (PCR) products has proven to be an effective alternative to the more traditional M13 sequencing technique. Advantages include the use of thermostable polymerases that allow high annealing and extension temperatures

(Gyllenstein and Erlich, *Proc. Natl. Acad. Sci. USA* 85:7652 (1988), Innis et al, *Proc. Natl. Acad. Sci. USA* 85:9436 (1988)), as well as the ability to directly utilize double-stranded DNA for templates (Carothers et al, *Biotechniques* 7:494; Sears et al, *Biotechniques* 13:626 (1992)). High temperature annealing and extension reduce ambiguous sequencing data that arise because of mispriming and/or secondary structure of the template. The capability of sequencing double-stranded DNA allows PCR products to be sequenced immediately after amplification. However, a simpler method for sequencing PCR products is to incorporate sequence delimiters directly into the PCR amplification process.

A sequencing method based on the incorporation of 5'- α -thiotriphosphates into PCR products has been demonstrated (Nakamaye et al, *Nucl. Acids Res.* 16:9947 (1988)). 5'- α -Thiotriphosphates were incorporated into DNA during PCR amplification and the positions of incorporated base-specific 5'- α -thiotriphosphates were revealed by chemical degradation with either 2-iodoethanol or 2,3-epoxy-1-propanol. Also 5'- α -thiotriphosphates have been used to sequence single stranded M13 DNA (Labeit et al, *Meth. Enzymol.* 155:166 (1987)). After incorporation by Klenow into primer extension products, the positions of the 5'- α -thiotriphosphates were revealed by exonuclease III

digestion. However, attempts to combine the best features of both methods (PCR amplification and enzymatic digestion) to reveal the sequence delimiters have proven unsatisfactory because of uneven band intensity (Nakamaye et al, *Nucl. Acids Res.* 16:9947 (1988); Olsen and Eckstein, *Nucl. Acids Res.* 17:9613 (1989)). The present invention overcomes the problems of the art and provides a simple and accurate method of amplifying and sequencing nucleic acids in a single step. Furthermore, the method is fast and amenable to automation.

OBJECTS AND SUMMARY OF THE INVENTION

It is a general object of the invention to provide a method of enzymatically incorporating into a nucleic acid a modified nucleotide which, once incorporated, is stable to chemical or enzymatic degradation. It is a specific object of the invention to provide methods of nucleic acid amplification, including strand displacement amplification, nucleic acid sequencing, *in vitro* transcription and gene therapy. It is a further object of the invention to provide methods of probing molecular interactions of nucleic acids with other cellular components, which methods are based on the enzymatic incorporation into the nucleic acids of

modified nucleotides, particularly, α -boronated deoxynucleoside triphosphates.

In one embodiment, the present invention relates to a method of synthesizing a nucleic acid fragment comprising:

contacting a nucleic acid template with:

- i) a primer sufficiently complementary to a portion of the template to hybridize therewith,
- ii) an enzyme that extends the primer so that a product complementary to the template is produced, and
- iii) four different nucleotides at least one of which, once incorporated into the nucleic acid, is resistant to enzymatic degradation,

the contacting being effected under conditions such that the at least one nucleotide is recognized by the enzyme and is thereby incorporated into the extension product of the primer.

In another embodiment, the present invention relates to a method of sequencing a nucleic acid comprising:

- i) enzymatically amplifying the nucleic acid in the presence of (a) four nucleotides that, once incorporated into a product of

the amplification, are susceptible to enzymatic degradation, and (b) a first modified nucleotide that is selectively incorporated into the product of the amplification in lieu of a first of the four nucleotides of (a), which first modified nucleotide, once incorporated into the product of the amplification is resistant to enzymatic degradation, and

repeating the enzymatic amplification in the presence of a second, third and fourth modified nucleotide, each of which second, third and fourth modified nucleotides is selectively incorporated into the product of the amplification in lieu of a second, third and fourth of the nucleotides of (a), each of the second, third and fourth modified nucleotides being resistant to enzymatic degradation once incorporated into the product of the amplification;

ii) treating the products of the amplification of step (i) containing the first, second, third and fourth modified nucleotides with an enzyme that digests

the products in the 3' to 5' direction,
the digestions terminating at the sites
of incorporation of the modified
nucleotides;

5 iii) separating the fragments resulting from
the treatment of step (ii) and detecting
the position of each of the modified
nucleotides in the products of the
amplification.

10 In a further embodiment, the present invention
relates to a method of sequencing a nucleic acid
comprising:

15 i) enzymatically amplifying the nucleic acid
in the presence of (a) four nucleotides
that, once incorporated into a product of
the amplification, are susceptible to
enzymatic degradation, and (b) a first
20 modified nucleotide that is selectively
incorporated into the product of the
amplification in lieu of a first of the
four nucleotides of (a), which first
modified nucleotide, once incorporated
into the product of the amplification is
resistant to enzymatic degradation, and

repeating the enzymatic amplification in the presence of a second, third and fourth modified nucleotide, each of which second, third and fourth modified nucleotides is selectively incorporated into the product of the amplification in lieu of a second, third and fourth of the nucleotides of (a), each of the second, third and fourth modified nucleotides being resistant to enzymatic degradation once incorporated into the product of the amplification;

- ii) subjecting the products of the amplification of step (i) containing a first, second, third and fourth modified nucleotides to mass spectrometry and detecting the resulting fragment patterns and thereby the position of each of the modified nucleotides in the products of the amplification.

In yet another embodiment, the present invention relates to a method of producing a protein comprising introducing into a cell a nucleic acid sequence encoding the protein, which nucleic acid sequence includes a modified nucleotide that is resistant to enzymatic degradation, the introduction being effected under

conditions such that the nucleic acid sequence is expressed and the protein thereby produced.

In yet another embodiment, the present invention relates to a method of amplifying a DNA sequence comprising:

5 i) contacting the DNA sequence with a primer having a first part and a second part,

the first part having a nucleotide sequence such that the first part hybridizes to a portion of the DNA sequence, and

10 the second part being 5' to the first part, noncomplementary to the DNA sequence and having a nucleotide sequence that includes a restriction enzyme recognition site,

15 wherein the contacting is effected under conditions such that the first part of the primer hybridizes to the DNA sequence; and

20 ii) enzymatically extending the primer and the DNA sequence in the presence of a boronated deoxynucleoside triphosphate so that a boronated duplex extension product is produced, the boronated deoxynucleoside triphosphate being selected so that a boronated deoxynucleoside is inserted into the extended DNA sequence at the site of cleavage of the restriction enzyme;

25

iii) contacting the duplex extension product with the restriction enzyme under conditions such that the second part of the primer is nicked; and

5 iv) contacting the nicked duplex extension product resulting from step (iii) with a polymerase that effects strand displacement under conditions such that a nucleic acid is produced from the site of the nick that is complementary to the extended DNA sequence.

10 Further objects and advantages of the present invention will be clear from the description that follows.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. One step PCR sequencing with α -boronated deoxynucleoside triphosphates (dNT^bP's) (Fig. 1A).
15 Bidirectional one-step sequencing with magnetic beads (Fig. 1B).

Figure 2. A portion of M13mp2 sequenced by the one-step process described in Figure 1A. Primer SS20 was labeled and used, along with primer RP, for PCR
20 amplification (250 bp product). Base-specific dNT^bP's were incorporated in four separate reactions and their positions were revealed by digestion with exonuclease III. The digested PCR products were separated by two

loadings on a denaturing polyacrylamide gel. As is shown in the first four lanes, the first loading separated fragments from position G₅₅ to about position A₂₁₀. The second loading separated fragments beginning at the first position beyond the 3' end of the primer, position G₂₁, to about position C₁₀₉. Missing bands are denoted by circled numbers and extra bands are noted in parentheses.

Figure 3. The sequence for M13mp2 extending from position G₂₁.

Figure 4. dNT^bPs as substrates for PCR.

PCR. PCR was performed in the presence of: all normal dNTPs, lane 1; normal dNTPs + 2% dAT^bP, lane 3; normal dNTPs + 2% dTT^bP, lane 5; normal dNTPs + 2% dGT^bP, lane 7; and normal dNTPs + 4% dCT^bP, lane 9.

Exonuclease. An aliquot of each PCR reaction was digested with exonuclease III: all normal dNTPs, lane 2; normal dNTPs + 2% dAT^bP, lane 4; normal dNTPs + 2% dTT^bP, lane 6; normal dNTPs + 2% dGT^bP, lane 8; and normal dNTPs + 4% dCT^bP, lane 10. DNA marker is in lanes labeled "M".

Figure 5. Primer extension/exonuclease III digestion with dNT^bPs.

Extension. The primer (unextended, lanes labeled P) was extended to the full length of the corresponding template in all instances: all normal dNTPs, lane 1; dAT^bP + normal dT-, dG-, dCTP, lane 3; dTT^bP + normal dA-, dG-, dCTP, lane 5; dGT^bP + normal dA-, dT-, dCTP, lane 7; and dCT^bP + normal dA-, dT-, dGTP, lane 9.

Exonuclease. Following extension, an aliquot of each sample was digested with exonuclease III. The all-normal product was digested to the limit of a duplex substrate, lane 2 (exonuclease III requires double-stranded substrate). In each case in which a boronated deoxynucleotide was substituted for a normal, exonuclease digestion was halted at the position of the boronated substitution: at A, lane 4; at T, lane 6; at G, lane 8; and at C, lane 10.

Figure 6. One-step PCR sequencing with end-labeled primer. Figure 6A. Both strands of a portion of phage T7 were sequenced. The top strand was sequenced by labeling primer T7_{rev}, likewise the bottom strand was sequenced by labelling primer T7_{for}. In each case, PCR amplification was accomplished with one labeled and one unlabeled primer (509 bp product). Base-specific dNT^bPs were incorporated in four separate reactions and their positions were revealed by digestion with exonuclease III. The forward sequence could be determined from C₃₄₅₈₆

to G₃₄₉₁₆; the reverse sequence could be determined from A₃₄₆₆₀ to G₃₅₀₀₀ (second loading not shown). Therefore, both strands were sequenced over approximately 350 bases. Figure 6B. One-step sequencing was repeated for the T7 samples as described for Figure 6A except that the amount of exonuclease III was increased from 25 units to 130 units. The sequence for phage T7 extending from position 34534 to 35042 is:

```

                                GGAGCG TAGGAAATAA
10  TACGACTCAC TATAGGGAGA GCGGAAATAA TCTTCTCCCT GTAGTCTCTT 600
    AGATTTACTT TAAGGAGGTC AAATGGCTAA CGTAATTAAA ACCGTTTGA
    CTTACCACTT AGATGGCTCC AATCGTGATT TTAATATCCC GTTTGAGTAT 700
    CTAGCCCGTA AGTTCGTAGT GGTAACCTTT ATTGGTGTAG ACCGAAAGGT
    CCTTACGATT AATACAGACT ATCGCTTTGC TACACGTACT ACTATCTCTC 800
15  TGACAAAGGC TTGGGGTCCA GCCGATGGCT ACACGACCAT CGAGTTACGT
    CGAGTAACCT CCACTACCGA CCGATTGGTT GACTTTACGG ATGGTTCAAT 900
    CCTCCGCGCG TATGACCTTA ACGTCGCTCA GATTCAAACG ATGCACGTAG
    CGGAAGAGGC CCGTGACCTC ACTACGGATA CTATCGGTGT CAATAACGAT 1000
    GGTCACTTGG ATGCTCGTGG TCGTCGAATT GTGAACCTAG CGA

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20 Figure 7. Bi-directional one-step PCR sequencing with a biotinylated primer. Both strands of a portion of phage T7 were sequenced. The top strand was sequenced using primer phi9_{rev}; likewise the bottom strand was sequenced using primer phi9_{for} which was

25 biotinylated prior to PCR amplification. Base-specific dNT^bPs were incorporated in four separate PCR reactions

(629 bp product) and their positions were revealed by digestion with exonuclease III. The forward sequence

could be determined from T₂₁₉₀₀ to A₂₂₁₀₀; the reverse sequence could be determined from G₂₂₃₅₀ to G₂₂₀₅₀.

5 Therefore, both strands were sequenced for 200-300 bases from a single loading. For comparison, cycle sequencing (right lanes) was performed with labeled primers according to the manufacturer's instructions (Perkin Elmer). The sequence for phage T7 extending from
10 position 21786 to 22414 is:

```

                                ACAG CTTACCTGA    800
GGCTATGGCT GCTGCCGCTG ATTCCGTAGG TTTACAGCCG GGAATTTAAT
ACGACTCACT ATAGGGAGAC CTCATCTTTG AAATGAGCGA TGACAAGAGG    900
TTGGAGTCCT CGGTCTTCCT GTAGTTCAAC TTTAAGGAGA CAATAATAAT
15 GGCTGAATCT AATGCAGACG TATATGCATC TTTTGGCGTG AACTCCGCTG    1000
TGATGTCTGG TGGTTCCGTT GAGGAACATG AGCAGAACAT GCTGGCTCTT
GATGTTGCTG CCCGTGATGG CGATGATGCA ATCGAGTTAG CGTCAGACGA    1100
AGTGGAACA GAACGTGACC TGTATGACAA CTCTGACCCG TTCGGTCAAG
AGGATGACGA AGGCCGCATT CAGGTTGTA TCGGTGATGG CTCTGAGCCG    1200
20 ACCGATGTGG ACACTGGAGA AGAAGGCGTT GAGGGCACCG AAGGTTCCGA
AGAGTTTACC CCACTGGGCG AGACTCCAGA AGAACTGGTA GCTGCCTCTG    1300
AGCAACTTGG TGAGCACGAA GAGGGCTTCC AAGAGATGAT TAACATTGCT
GCTGAGCGTG GCATGAGTGT CGAGACCATT GAGGCTATCC AGCGTGAGTA    1400
CGAGGAGAAC GAAGA

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Figure 8. PCR amplification using boronated dNTPs and in vitro transcription of boronated templates.

Upper panel. PCR amplification. The reaction mixture (50 μ l total) contained T7 DNA (10 μ g), forward primer (T7 positions 34534-34553; 10 pmol), reverse primer (complementary to T7 positions 34625-34644), PCR buffer, Vent DNA polymerase (New England Biolabs, 1 unit), and dNTPs (200 μ M) which were the normal triphosphates except that boronated triphosphates were substituted for the normal one(s) as follows: all normal (lane 2), boronated dGTP (lane 3), boronated dCTP (lane 4), boronated dATP (lane 5), boronated dGTP+dCTP (lane 6), boronated dGTP+dATP (lane 7), boronated dCTP+dATP (lane 8), and boronated dGTP+dCTP+dATP (lane 9). The amplification was performed for 25 cycles of 95°C for 1', 53°C for 1', and 76°C for 1'.

Lower panel: In vitro transcription. The reaction mixture (55 μ l total) contained DNA from the PCR reactions (10 μ l), translation buffer (final concentrations: 40 mM Tris-HCl (pH 8.1), 20 mM MgCl₂, 1 mM spermidine, 5 mM DTT, 5 μ g/mL BSA, and 0.01% Triton X-100; 11.3 μ l), RNasin (USB, 3 μ l), T7 RNA polymerase (80 units), and NTPs (2 mM each). The reaction was carried out for 2 hours at 37°C. The substitution pattern of boronated templates corresponds to that described for the upper panel, ie all normal (lane 2),

boronated dGTP (lane 3), boronated dCTP (lane 4),
boronated dATP (lane 5), boronated dGTP+dCTP (lane 6),
boronated dGTP+dATP (lane 7), boronated dCTP+dATP (lane
8), and boronated dGTP+dCTP+dATP (lane 9).

5 Electrophoresis. All samples were mixed with a
sucrose loading buffer, separated on a 5% agarose gel,
and visualized by ethidium bromide fluorescence. Lane 1
of each panel contains marker DNA.

Figure 9. Incorporation of boronated dNTPs into
10 DNA by PCR; resistance of boronated DNA to mouse serum
and exonuclease digestion; transcription of boronated
DNA into mRNA; and translation of mRNA into protein.
Figure 9A. Mouse serum digestion of normal (lane 1) and
resistant boronated (lane 4) PCR products. Fig. 9B.
15 Production of PCR products - normal (lane 1), boronated
(lanes 2 and 3); exonuclease III digested normal PCR
products (lane 4) and resistant boronated PCR product
(lanes 5 and 6); transcription of normal and boronated
DNA (lanes 7-9). Fig. 9C. Translation of normal
20 (lane 1) and boronated (lane 2 dG^b, lane 3 dC^b) PCR
products.

Figure 10. Expression of normal and boronated
templates.

Figure 11. Resistance to 5' exonuclease digestion by boronated PCR products. Lane 1 - all normal, lane 2 - all normal/exo, lane 3 - dAT^bP, lane 4 - dAT^bP/exo, lane 5 - dTT^bP, lane 6 - dTT^bP/exo, lane 7 - dGT^bP, lane 8 - dGT^bP/exo, lane 9 - dCT^bP, lane 10 - dCT^bP/exo.

Note: The exo+ samples (lanes 2, 4, 6, 8, 10) appear to migrate more slowly than the exo- samples (lanes 1, 3, 5, 7, 9) because they were loaded at different times. The size of the fragments is the same.

Figure 12. Chemical synthesis of oligodeoxynucleotides containing internucleotide boranophosphate diester linkages.

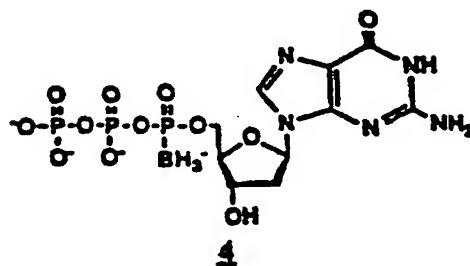
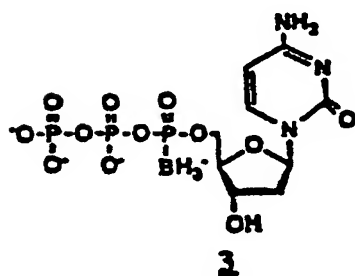
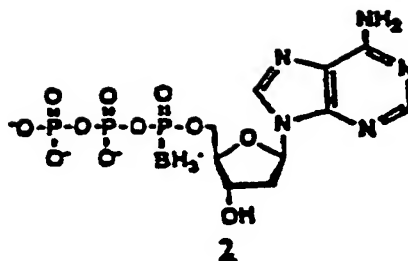
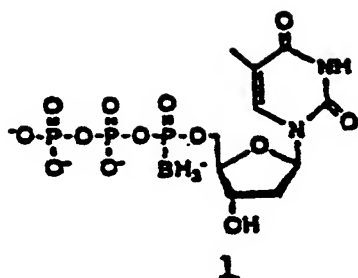
Figure 13. Sequence of a DNA duplex containing multiple restriction enzyme recognition sites.

DETAILED DESCRIPTION OF THE INVENTION

The present invention results, at least in part, from the discoveries that deoxynucleoside boranophosphates can be enzymatically incorporated into nucleic acids and that once incorporated, such nucleotides are stable to enzymatic (eg exonuclease) digestion. These discoveries make possible unique approaches to nucleic acid amplification and sequencing,

DNA structure determination and therapeutics, including antisense and gene therapy, and diagnostic approaches based, for example, on enzyme inhibition. The present invention relates to these approaches, as well as to
5 others.

In one embodiment, the present invention relates to a method of simultaneously amplifying a nucleic acid (eg, by PCR for DNA or by reverse transcriptase type reaction for RNA) and sequencing the product obtained.
10 The method involves the utilization of modified nucleotides that can be enzymatically incorporated into nucleic acids in lieu of their unmodified, naturally occurring counterparts. The modified nucleotides are, preferably, nucleoside borano-triphosphates, eg, 5'- α -
15 borano-triphosphates:



The borano moiety can be $-BH_2$ (as indicated above), or other such moiety as described in USP 5,177,198. The Examples set forth herein relate to boronated compounds, however, it will be appreciated that other elements

5 having a large neutron capture cross-section for low energy neutrons, and suitable for use in neutron capture therapy, such as gallium, can also be used. The base moiety of modified nucleotides suitable for use in the present invention can be a naturally occurring base or a

10 derivative, such as 7-deaza-guanine, inosine, 5-methylcytosine, etc (see USP 5,177,198). The use of modified bases can be advantageous for reasons noted in Current Protocols in Molecular Biology, pp. 7.0.1-

7.7.31, eds. Ausubel et al, Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (1993). One skilled in the art will appreciate that it may also be advantageous to utilize modified sugar moieties.

5 The base-specific incorporation of modified nucleotides (eg, 2'-deoxynucleoside 5'-borano-triphosphates) into DNA can be effected during primer extension using protocols such as PCR (described in USP 4,683,195 and 4,683,202). Other enzymatic protocols can
10 also be used, such as those employing diphosphates rather than triphosphates as substrates. The optimum polymerase incorporates dNTPs efficiently, base-specifically and independent of the surrounding sequence context. *Taq* polymerase and *Vent* polymerase can be
15 used, as can other thermostable polymerases. When RNA sequencing is sought, *Tth* polymerase can be used. Once incorporated, the modified nucleotides block the action of exonucleases, for example, exonuclease III, or other 3'-5' exonuclease that cleaves normal phosphodiester
20 but to which the modified nucleotide (eg, borano-phosphate) proves resistant, thereby making it possible to determine the position of the 3' borano-phosphate in each amplification product.

25 As an alternative to the exonuclease treatment described above, one skilled in the art will appreciate that techniques such as mass spectrometry can also be

used to detect the location of the modified nucleotide.

It has been shown that DNA can be sequenced by Matrix-Assisted Laser Desorption/Ionization Mass Spectrometry (MALDI-MS; Tang et al, *Rapid Comm. in Mass. Spectrom.* 7:63 (1993); Wu et al, *Rapid Comm. in Mass. Spectrom.* 7:142 (1993)). The procedure involves using mass spectrometry, which can be accomplished in minutes, to analyze sequencing reactions. With MALDI-MS, it has been possible to determine the sequence for about 40 nucleotides of standard base-specific sequencing reactions. Boron is found only in trace amounts in DNA or RNA and it has several natural isotopes. Therefore it is a good biomarker for mass spectrometry. Since boronated triphosphates can be incorporated base-specifically in sequencing reactions, it should be possible to attach a boron tag to the sequencing reaction products to aid in the separation and identification of the molecular fragments by MALDI-MS.

The method of the present invention, as it relates to one-step PCR sequencing using α boronated dNTP's, is depicted in Figure 1A. As shown in Figure 1A, the method can be carried out in four simple manipulations in a "one-pot" reaction:

1. Primer labeling: One of two PCR primers, specific for the region to be amplified, is labeled at

the 5' end with a detectable label, for example, a radioactive, fluorescent or chemiluminescent label. If both strands are to be sequenced, uniquely labeled primers for both strands must be provided.

5 (Alternatively (if bi-directional amplification is not required), the modified nucleotide to be incorporated can be labeled rather than the primer. The label can be on the base moiety, sugar moiety, or on the boron moiety of the modified nucleotide. In addition to radioactive,
10 fluorescent and chemiluminescent labels, binding pair members can also be used, such as biotin, as can other labels known in the art. Antibodies directed to moieties of the modified nucleotide can also be used for purposes of detection.)

15 2. Base-specific reactions: Four separate PCR amplifications are performed in the presence of the four normal dNTP's (dA, dT, dG and dC), plus one of the dNTP's, preferably, present at a low level (eg, for dA, 10:1 normal to boronated; for dT, 5:1, normal to
20 boronated; for dG, 10:1 normal to boronated; for dC, 1:2 normal to boronated). (Alternatively, the amplifications can be performed simultaneously using four polymerases selected so as to be specific for only one of the boronated dNTP's.)

3. Exonuclease digestion: Each of the base-specific PCR reaction products is digested with 3' to 5' exonuclease, for example, exonuclease III, to produce fragments that terminate at base-specific dNM^bPs.

5 (Alternatively, fragmentation can be effected, as well as detection, using the spectrometric method described above.)

4. Fragment separation: Upon separation (eg, by electrophoresis), the borano-terminated fragments
10 produce a uniform band pattern marking the position of each base in the PCR product.

The above-described process is advantageous as it is direct - the primers can be pre-labeled and the sequence delimiters (eg, the boranophosphates) can be
15 incorporated during amplification. Therefore, the present method eliminates the need to remove unextended primers and truncated extension products from the PCR products in preparation for sequencing. Further, the
20 nucleic acid purification and the secondary cycle sequencing procedure can be replaced by a simple and completely automatable exonuclease digestion that reveals the chain truncators (the modified nucleotides). The one-step solution to cycle sequencing afforded by the present method reduces the time substantially

between initiation of amplification and separation of the fragments obtained (it should be noted, however, that borano-containing compounds can also be used as truncators in a cycle sequencing approach). Just as
5 important as the time savings is the elimination of the nucleic acid purification step which renders the technique amenable to automation, as described below.

In addition to the protocol depicted in Figure 1A, the present method can also be used to effect
10 bidirectional sequencing. An exemplary approach utilizing magnetic beads is set forth in Figure 1B and is described below. Magnetic bead technology has been demonstrated to be an effective method for the dideoxy sequencing of PCR products (Hultman et al, Nucl. Acids
15 Res. 17:4937 (1989), Biotechniques 10:84 (1991)). In the context of the present method, magnetic beads can: (1) facilitate exonuclease digestion of the PCR products and (2) allow both strands of the PCR template to be sequenced. First, one of the primers is modified with a
20 first member of a binding pair, for example, biotin and then a label and boronated nucleotides are incorporated during amplification. Following amplification, the duplex DNA is immobilized to magnetic beads linked to the other member of the binding pair (eg when biotin is
25 the first member of the binding pair, streptavidin-linked magnetic beads can be used) and treated with

exonuclease. Following digestion, the fragments from each strand are isolated independently and separated, for example, by PAGE to produce the sequence. The procedure shown in Figure 1B is outlined below.

5 1. Perform four base-specific dA, dT, dG, and dC reactions: Four separate PCR amplifications are performed with one biotinylated and one unmodified primer in the presence of all four normal dNTPs plus a few percent of one of the dNTPs (eg, for dA, 10:1
10 normal to boronated; for dT, 5:1, normal to boronated; for dG, 10:1 normal to boronated; for dC, 1:2 normal to boronated). Also, sufficient labelled dNTP (radioactive or other) is added to the reaction such that the boronated chain delimiter (and label, if appropriate) is
15 incorporated during PCR amplification.

 2. Bind the PCR products to streptavidin-linked magnetic beads. The biotinylated PCR products are incubated with streptavidin-linked magnetic beads and then the PCR product-biotin-streptavidin-magnetic bead
20 complexes can be immobilized with a magnet. While the complexes are immobilized, they can be washed to remove PCR reactants and the buffer changed to the optimal exonuclease buffer.

3. Digest with exonuclease III. While still attached to the magnetic beads, each of the base-specific PCR reaction products is digested with exonuclease III to produce fragments which terminate at base-specific dNM^bPs.

4. Remove the unmodified primer strand. Digested fragments derived from the unmodified (eg, non-biotinylated) primer strand are separated from the biotinylated strand, for example, by heating the digestion reaction mixture. The biotinylated strand fragments are immobilized with the magnet and the unmodified fragments are removed in the exonuclease buffer and loaded onto the sequencing gel.

5. Remove the biotinylated strand from the magnetic beads. Digested fragments derived from the biotinylated primer can be removed from the streptavidin-linked magnetic beads, for example, by heating the complexes in loading buffer (see Tong et al, Anal. Chem. 64:2672 (1992)). The free magnetic beads are immobilized with the magnet and the biotinylated fragments are removed in loading buffer and loaded directly onto the sequencing gel.

While the foregoing is described by way of reference to a biotin/avidin binding pair, alternative binding pairs can also be used.

5 In addition to the approaches described in Figures 1A and 1B, the present invention relates to a method of "mirror-image" sequencing. In this method, 5' and 3' exonucleases are used in concert to produce mirror image sequence data derived from each strand of duplex DNA. For example, a duplex PCR product is digested from the
10 same end by using a 5' exonuclease, such as T7 gene 6 or λ exonuclease, for the Watson strand, and a 3' exonuclease for the Crick strand. In each case, digestion proceeds until the nuclease encounters a base-specific boronated nucleotide, at which point digestion
15 is blocked, thereby producing a family of sequencing fragments. Since the fragments derived from the two complementary strands are digested from the same end, the length of the fragments at each position in the sequence will be the same. Therefore the sequencing gel
20 will produce side-by-side mirror images of the sequence, thus providing immediate and independent sequence verification.

"Mirror-image" sequencing involves the use of a 5'-exonuclease that shows differential digestion towards
25 normal and borano-phosphates. The degree of susceptibility of the borano-phosphates to an

exonuclease can be determined experimentally. Using an appropriate pair of nucleases and borano-stereoisomers, the experiment is performed as follows: (a) following PCR, the labeled products are split into separate tubes and digested with a double-strand specific 5' or 3' exonuclease, generating two sequencing ladders, one each from the Watson and Crick strands; or (b) one biotinylated primer is used and the label and boranophosphates are incorporated during PCR. The strands are then separated into two vessels where the 5' and 3' exonuclease digestions are performed. Nucleases here must be single-strand specific, such as exonuclease I (3') and phosphodiesterase II (5').

The potential uses for the sequencing procedures described herein will be clear to those skilled in the art. For example:

(i) The shotgun method of large scale sequencing relies on breaking up cosmid-sized DNA into smaller pieces that are cloned into phage M13. The ssM13 templates are then sequenced with universal primers. Conventionally, the templates are amplified by PCR or bacterial culture, purified, and sequenced. The method of the present invention makes it possible to pick a virally infected bacterial plaque or cell, amplify the template with a labeled primer in the presence of dNT^bPs, and, after exonuclease digestion, load the

samples onto a gel. The time savings is substantial because the purification and cycle sequencing steps are replaced with a simple exonuclease digestion.

5 (ii) Alu PCR takes advantage of the ubiquitous Alu repeat sequence in human DNA to design PCR primers that amplify human DNA (Nelson et al, *Proc. Natl. Acad. Sci. USA* 86:6686 (1989)). The technique has been useful for amplifying inter-Alu sequences from somatic cell hybrids and from flow-sorted chromosomes (Nelson et al, *Proc.*
10 *Natl. Acad. Sci. USA* 88:6157 (1991); Cotter et al, *Genomics* 9:473 (1991)). The one-step procedure of the invention makes it possible to amplify Alu PCR products that can be sequenced or cloned directly. That is, the products can be sequenced by the usual exonuclease III
15 digestion method, or, by simply omitting the digestion, the PCR products can be cloned.

(iii) One focus of the human genome project is a global survey of human genetic diversity (Cavalli-Sforza et al, *Genomics* 11:490 (1991); Baer, *Human Biology* 65:7
20 (1993)). Direct sequencing of PCR products makes it possible to perform high-resolution analysis of many individuals from a variety of geographic locations (Ritte et al, *Human Biology* 65:359 (1993)). For
25 example, mitochondrial DNA, amplified from single plucked human hairs, has been used to determine the population diversity among individuals of an aboriginal

African community (Vigilant et al, *Proc. Natl. Acad. Sci. USA* 86:9350 (1989)). The one-step procedure of the invention is ideally suited for such studies because:

(1) few primers are required because only a few selected genomic markers are chosen to be amplified, and (2) template DNA is virtually unlimited because of the ease of collection and stability of human hair. Therefore, the rate of population diversity data collection can be increased by the time-savings method of the invention.

(iv) The procedure of the invention can also be applied to disease diagnosis. For example the majority of individuals afflicted with cystic fibrosis carry a characteristic three-base deletion, termed ΔF_{508} (Riordon et al, *Science* 245:1066 (1989); Bat-sheva et al, *Science* 245:1073 (1989)). Individuals can be tested for the ΔF_{508} deletion by analysis of heteroduplex DNA (Rommens et al, *Am. J. Hum. Genet.* 46L395 (1990)). However, some individuals have the cystic fibrosis phenotype but do not carry the ΔF_{508} deletion; thus the genotype of these individuals must be determined by sequencing (Shoshani et al, *Genomics* 15:236 (1993); Lucotte and Loirat, *Meth. Enzymol.* 155:166 (1993)). The present method can accelerate the rate of genotyping of individuals that are either diseased or are carriers, but that do not show the ΔF_{508} deletion. Many other diseases that can be diagnosed by sequencing would benefit from the time-

savings of the present method. Likewise, the present invention can be applied in forensics, and evolutionary biology.

5 In contrast to methodologies of the art, the entire one-step PCR sequencing procedure of the invention is amenable to automation. The procedure requires only four processes (as described above) because the amplification and termination steps are accomplished simultaneously: (1) select templates, (2) amplify
10 incorporating the sequence-delimiting modified dNTP's, (3) digest with exonuclease to unmask the modified dNMP's, and (4) separate and detect the DNA fragments. Processes 2 and 3 are "one pot" reactions; therefore, no human intervention is necessary beyond selecting the
15 template and providing a robotic workstation with reagents. One skilled in the art will appreciate that the need for four separate amplification reactions can be avoided by selecting polymerases, produced, for example, by site directed mutagenesis, that are specific
20 for one of the bases present in the nucleotide to be incorporated and including each such polymerases in the same reaction vessel.

The ABI CATALYST 800 Molecular Biology LabStation, for example, is well-suited to perform the present
25 method automatically. It is designed specifically to perform sequencing reactions and is pre-programmed to

perform cycle sequencing (some modification of the CATALYST software would be required).

5 Upon completion of the exonuclease step of the present procedure, the samples are, advantageously, loaded onto a gel (the Bio-Rad Gene Loader II, for example, can load gels automatically). If radioactive or chemiluminescent labels are used, one sample is, advantageously, loaded per lane. The advantage of a radioactive label is the simplicity of 5' end-labeling the primer by polynucleotide kinase. Alternatively, a chemiluminescently-labeled primer (or other non-radioactively labeled primer) makes it possible to avoid the contamination and disposal problems associated with radioactively-labeled samples. Additionally, the present one-step sequencing protocol can be expected to be compatible with multiplex sequencing in which labeling is accomplished, after transfer of the gel to a solid support, by hybridization to a radioactively- or chemiluminescently-labeled probe (Church and Gilbert, *Proc. Natl. Acad. Sci. USA* 81:1991 (1984); Church and Kieffer-Higgins, *Science* 240:185 (1989)).

Fluorescent labeling options are:

(1) incorporation of fluorescently-labeled chain termination during polymerization (Dye-deoxy terminators, ABI) or (2) utilization of pre-labeled fluorescently-labeled primers. Since the present

procedure requires that the 3' ends of the product molecules be removed by exonuclease, the fluorescent labels are, advantageously, attached to the primers prior to amplification. In the simplest case, one primer is labeled per base-specific reaction and the G, C, T, and A samples are loaded in separate lanes. However, the ABI373A sequencer, for example, is capable of sequencing all four bases in a single lane, and the present method is adaptable to one-lane sequencing.

Fluorescently-labeled boronated triphosphates can serve both as label and sequence delimiters in the present processes. Synthesis of such triphosphates can be carried out as follows. The first step is the attachment of the 3-amino-1-propyn-1-yl linker arm to the 5 position following the procedure of Robins et al. The 5'-phosphate residues are then attached in three successive steps following Tomasz et al (Angew. Chem. Int. Ed. Engl. 31:1373 (1992)). The crucial step is the treatment with pyridine-borane to effect boronation of the phosphate; however, as a result of the low reactivity of pyridine-borane, reduction of the propynyl moiety of the linker is highly improbable. Slightly alkaline treatment of the Lewis-base borane formed in this reaction will result not only in the elimination of the β -cyanoethyl group from the phosphorous, but also in selective 3'-O-deacetylation. After building-up the

5'-boronated-triphosphate residue, the N-trifluoroacetyl protecting group will be removed from the linker. The diastereomers can be separated either before or after this step. Finally, the peptide bond is formed between the carboxyl group of the appropriate succinimidylfluorescein dye (Applied Biosystems) and the amino group of the linker, followed by removal of the acetyl protecting group from the phenolic hydroxyl function of the dye. (Alternative linkers can be designed (eg a n-carbon alkyl chain) using standard chemistries (see Zaratova et al, Nucleosides and Nucleotides 10:295 (1991) and Vincent et al, Nucleic Acids Res. 10:6787 (1982)).

In order to employ one-lane sequencing with the present method, the PCR primers are labeled with base-specific (eg, fluorescent) tags. For example four fluorescent phosphoramidite labels can be obtained from ABI that can be attached to the 5' end of oligonucleotides during solid phase synthesis; therefore, all four bases can be sequenced in a single lane. Further, because in the present method PCR amplification and incorporation of sequence delimiters are accomplished simultaneously, the method has the potential to sequence, bi-directionally, both strands of the PCR product in a single lane.

The rationale for bi-directional one-lane sequencing is as follows. Each position of a sequencing gel ladder can be thought of as an array of binary data in which either a band is present (1) or absent (0)

5 (Nelson et al, Nucl. Acids. Res. 20:1345 (1992)). A four lane, one label sequencing gel (or likewise, a one lane, four label gel) represents each base by the presence of a band at each position, for example: A = (1,0,0,0), T = (0,1,0,0), G = (0,0,1,0), C = (0,0,0,1).
10 However, a binary system with four lanes (or labels) is capable of determining $2^n - 1$ outcomes (to avoid the null outcome). Therefore, the four bases of each strand can be represented by a unique combination of four labels, for example: $A_1 = (1,0,0,0)$, $T_1 = (0,1,0,0)$, $G_1 =$
15 $(0,0,1,0)$, $C_1 = (0,0,0,1)$, and $A_2 = (1,1,0,0)$, $T_2 = (0,1,1,0)$, $G_2 = (0,0,1,1)$, $C_2 = (1,0,0,1)$. Consequently, the two primers can be labeled with a combination of the four different fluorescent labels and used separately in base-specific reactions that, after pooling, make it
20 possible to sequence all four bases of both strands of the DNA template in a single lane.

In order for the reaction products of the invention to be loaded directly onto a gel, a relatively high concentration of labeled primer is advantageously
25 present during amplification. The results shown in Figure 2 were obtained from a 25 μ l reaction that was

concentrated by precipitation prior to loading. It is noted that ABI has developed a low-volume method for fluorescent sequencing (Prism). In their method, the total amount of primer is held constant but the concentration is increased from 0.4 pmol/ μ l to 1.2 pmol/ μ l, thus allowing sequencing reactions to be pooled, mixed with loading buffer, and loaded directly onto a gel. It is noted that 0.4 pmol/ μ l is the same concentration used the presently described protocol. Therefore, increasing the primer concentration of the present protocol by a factor of 3, while reducing the volume to 5-10 μ l, should allow the reactions to be loaded directly onto a sequencing gel.

In any series of PCR amplification experiments, carryover of PCR products from prior experiments is a concern. The most effective prevention of contamination is to separate physically the PCR reactants from the products (Kwok, *PCR Protocols: A guide to Methods and Applications*, pp. 142-145, Academic Press (1990)). However, physical separation is not possible when using the CATALYST 800 robot because both the pipet tip and the reaction wells are reusable. Since the present invention includes PCR amplification with sequencing, the procedure will be sensitive to contamination by PCR products. Therefore, after each experiment, the pipet tip and reaction wells could be removed from the

CATALYST and washed thoroughly or decontaminated by some other method.

In addition to the one step procedures described above, boronated triphosphates can also be employed to extend the usefulness of strand displacement amplification (SDA) (Walker et al Proc. Natl. Acad. Sci. USA 89:329-396). SDA accomplishes in vitro DNA amplification by utilizing specific properties of 2'-deoxyadenosine 5'- α -thio-triphosphate, ie, it can be incorporated into DNA by exonuclease-free Klenow and, once incorporated, it is resistant to digestion by the restriction enzyme *HincII*. The resistance to digestion results in a primer extension product which is nicked at the synthetic primer site, but unaffected at the sites of 2'-deoxyadenosine 5'- α -thio-triphosphate incorporation; thus the protected strand becomes available to serve as the template for subsequent cycles of amplification. However, the applicability of SDA is limited because only relatively short DNA fragments (<100 bp) can be amplified and the procedure exhibits a sequence specificity which prevents amplification of certain regions of DNA. The limitations of SDA are probably due, in part, to the relatively poor incorporation efficiency of 2'-deoxyadenosine 5'- α -thio-triphosphate. Since the boronated compounds are resistant to nucleases and are very good substrates for

exonuclease-free Klenow, ie the efficiencies of
incorporation by exonuclease-free Klenow (V_{max}/K_m) are
comparable to their natural counterparts, both the
length of amplified fragments and the variety of target
5 sequences can be expected to be increased by
substituting boronated triphosphates directly into the
SDA procedure.

Boranophosphates can also be used in long range PCR
sequencing and closure. The conventional methods for
10 long range sequencing and closure are primer walking and
the cloning of sets of unidirectional nested deletions.
Boranophosphates provide an alternative to these
techniques. It has been shown recently that very long
DNA duplexes can be produced by PCR (Barnes, Proc. Natl.
15 Acad. Sci. USA 91:2216 (1994)); both an *exo+* and an *exo-*
thermostable polymerase were used in the PCR reaction.
(*Taq* (*exo-*) and Deep Vent (*exo+*) have been used to
amplify efficiently a 13-kb region from phage T7 DNA.
Amplification was achieved in high yield with all normal
20 dNTPs as well as with successive addition of the
boronated dNTPs (2.5 μ M).) Since a maximum of 1-kb of
DNA can be sequenced by the best of sequencing runs,
both strands of a duplex of a greater size could not be
sequenced directly by conventional techniques. However,
25 the use of boronated dNTPs in the amplification provides

a means of sequencing without the need for an intermediate primer walking or nested deletion step.

In accordance with this embodiment of the present invention, a long amplified duplex is fragmented using a restriction enzyme(s) and the fragments separated, for example, by agarose gel electrophoresis. The restriction fragments can be, for example, cut out of the gel and labeled by back-filling at a 5' overhang. After labeling (eg, at the 3' end), the boronated sequence delimiters are revealed by a 5' exonuclease that acts on double stranded DNA, such as T7 gene 6- or λ - exonuclease. The boranophosphates are resistant to these 5' exonucleases, as shown in Example 9 (see also Thatcher et al, J. Org. Chem. 58:2272 (1993)). Thus, once the long DNA duplexes are amplified, restricted into smaller fragments, labeled by back-filling, and digested by exonuclease, the fragments can be, for example, loaded directly onto a polyacrylamide gel and sequenced. The number of manipulations involved in this procedure is about the same as needed for current methods for preparing sets of nested primers and cloning, however, in the present case, the sequence of the DNA is obtained. It will be appreciated that, in addition to using double-strand specific 5' exonucleases, the strands can be denatured and then single-strand specific exonucleases (eg

phosphodiesterase II) used. It is also noted that boranophosphates provide a ready way to produce a set of nested deletions for cloning. Random incorporation of boranophosphates during PCR followed by exonuclease III provides nested sets of deletions that terminate at a boranophosphate. These fragments can be ligated into cloning vectors. The lengths of the fragments can be determined statistically by the percentage of boron doping.

Boronated-triphosphates (ribo or deoxyribo) can also be used in the preparation of aptamers. Aptamers are oligonucleotide ligands selected from a combinatorial "shape" library to fit a target (which can be a small molecule like caffeine or theobromine, or a large molecule like a protein, oligosaccharide, oligonucleotide, enzyme, antibody, receptor or other regulatory molecule (see Kenan et al, TIBS 19:57 (1994); Szostak TIBS 17:89 (1992); Green et al, Science 258:1910 (1992); Wang et al, Biochemistry 32:1899 (1993); Tuerk et al, Science 249:505 (1990); Beaudry et al, Science 257:635 (1992)). DNA and RNA are capable of mimicking the shapes of proteins and other molecules, and their ability to be amplified by PCR makes it possible to select one molecule from a population of 10^{15} molecules prepared by production of random combinations of building blocks at every position of a macromolecule.

However, a limitation of nucleic acids is that there are only 4 naturally occurring monomer units from which to assemble complex oligomeric ligands. Since boronated triphosphates are good substrates for PCR, they are
5 suitable for aptamer assembly, thereby providing 4 additional building blocks. Therefore, boronated triphosphates (of both RNA and DNA, as well as with modified bases) increase the variety of ligands that can be produced by aptamer technology.

10 In accordance with this embodiment of the present invention, boronated triphosphates (ribo or deoxyribo) are used as monomer units for production of a nucleic acid (DNA or RNA) shape library by PCR. The library is designed to have degeneracy at a number of different
15 positions in the oligonucleotide. The combinatorial library is screened for selection against a target surface. Selected ligands that bind to the target are partitioned from those unbound, producing a population of molecules that is enriched in binding affinity to the
20 target. The enriched population is amplified by PCR. By subjecting the enriched population to further rounds of binding, higher affinity ligands are progressively selected. These progressive cycles of amplification and reselection produce optimized ligands, or aptamers.

25 In addition to amplification/sequencing protocols, nucleic acids containing modified nucleotides of the

type described herein, advantageously boronated nucleic acids, can be used to probe the interaction of nucleic acids with other cellular components. One skilled in the art will appreciate that nucleic acids containing modified nucleotides (eg boronated nucleotides) can be used in electron spectroscopic imaging (Bendayan et al, *J. of Histochem Cytochem* 37:573 (1989)) and electron energy loss spectroscopy (Colliex, *Ann. NY Acad. Sci.* 483:311 (1986); Ottensmeyer, *J. Ultrastruct. Res.* 88:121 (1984) and 72:336 (1980)).

Further to the above, α -borano triphosphates can be expected to be useful for direct gene transfer regimens. The properties of α -boronated triphosphates that can be utilized to sequence DNA, ie the compounds can be incorporated into DNA by polymerases and, once incorporated, are resistant to nucleases, can also be used in direct gene transfer. In direct gene transfer, DNA that includes a promoter, a gene of interest, and a terminator is injected, for example, into muscle tissue (Wolff et al, *Science* 247:1465-1468 (1990)). The introduced genes can be transcribed into RNA and translated into proteins that can perform a variety of pharmacological functions. For example, such proteins have been shown to: modulate the hormonal levels of cardiac tissue in rats (Kitsis et al, *Proc. Natl. Acad. Sci.* 88:4138-4142 (1991)), express foreign genes in fish

(Hansen et al, *FEBS Letters* 290:73-76 (1991)), express human dystrophin in mice (Acsadi et al, *Nature* 352:815-818 (1991)), elicit an immune response to a foreign protein in mice (Tang et al, *Nature* 356:152-154 (1992)), and elicit protection against the influenza virus in mice (Ulmer et al, *Science* 259:1745-1748 (1993)). Expression of the injected genes can be detected over long periods of time, ie several months, even though the DNA is degraded rapidly in blood serum. Since α -borano trisphosphates can be incorporated into DNA by polymerases, the stability that they exhibit towards nucleases should increase the lifetime of the injected DNA and thereby increase the efficacy of the direct gene transfer method.

It will be clear from the foregoing that synthesis of oligonucleotides (eg nucleic acids containing 2 or more nucleotide units) containing boranophosphate internucleotide linkages can be effected enzymatically (see also Example 11). Chemical synthesis's can also be effected, for example, to provide greater amounts of material. Previously, the phosphoramidite approach was used for synthesis of the boranophosphate oligonucleotides. The triester phosphite group formed during coupling reaction easily reacts with various borane complexes giving the desired boranophosphate. However, this methodology requires the boronation

reaction on each elongation cycle and therefore a high yield of this reaction is very critical for successful synthesis of long oligonucleotide chains. Close to quantitative yields can be achieved by using active
5 borane complexes such as THF:BH₃ or Me₂S:BH₃, but the probability of accompanying base damage significantly increases. An additional problem is that the overall yield of the desired product after addition of a third or successive nucleotide unit can be reduced, possibly
10 reflecting some undesirable reaction involving the boranophosphate group. The present invention provides an improved method for synthesis of boranophosphate oligonucleotides utilizing an H-phosphonate approach. In so doing, the present method avoids multiple
15 boronation steps and permits one boronation reaction for all H-phosphonate groups simultaneously after the complete chain elongation of an oligonucleotide.

More specifically, the present invention relates to an effective method of conversion of internucleotide
20 H-phosphonate diester groups to boranophosphate ones. In accordance with the present method, a H-phosphonate diester group is converted to a phosphite triester group. The phosphite triester group contains a free electron pair and is significantly more reactive towards
25 electron-accepting groups than the H-phosphonate diester. The boronation reaction of the phosphite

triester group can be done in mild conditions thereby reducing, if not essentially eliminating, base damage.

While the present method relates to the synthesis of oligonucleotides of any length, it is exemplified by reference to two model conversions of the H-phosphonate to the boranophosphate: dimethyl H-phosphonate and 5'-O-DMT-2-deoxythymidylyl[3'-5'] (3'-acetylthymidylyl) H-phosphonate (see Example 10). The course of the reactions has been followed by ^{31}P -NMR spectroscopy. The

first conversion of H-phosphonate diester to the phosphite triester has been achieved by silylation with bis(trimethylsilyl)acetamide. During the reaction, utilizing anhydrous conditions and inert atmosphere, the signals of H-phosphonate dinucleoside stereoisomers at $\delta=9.4$ and 11.2 ppm disappeared and new signals at $\delta=129.5$ and 130.0 ppm corresponding to the phosphite triester stereoisomers appeared. After complete disappearance of H-phosphonate signals and without separation of intermediate product, a large excess of pyridine-borane complex was added to the reaction mixture. Pyridine-borane complex does not significantly react with any of the four DNA bases under the conditions used for phosphite boronation. The boronation reaction leads to disappearance of the phosphite triester signals and appearance of a broad signal at $\delta = 104.9$ ppm corresponding to the

boranophosphate triester. Deprotection by ammoniacal treatment of the reaction mixture gives the desired boranophosphate diester signal at $\delta=94.3$ ppm. Overall yield of the conversion based on ^{31}P -NMR spectroscopy data is 60% for the dinucleoside H-phosphonate and 85% for dimethyl H-phosphonate. Changing the 5' -OH protecting group from DMT to O-trimethylsilyl can increase the yield by 10% or more.

While the present method can be carried out in solution, solid-phase synthesis can be more suitable for the synthesis of long oligonucleotides. Any of a variety of solid supports can be used to convert immobilized H-phosphonate to boranophosphate, silica gel support being preferred. Chemical procedures for solid-phase synthesis are essentially the same as for solution synthesis. Both silica gel support and polystyrene resin have been shown to result in high yields of final product (eg dithymidine boranophosphate), 75% and 60%, respectively, using manual synthesis. Yields approaching 100% can be achieved using an automated synthesizer because such systems make possible an anhydrous and anoxic environment.

Certain aspects of the present invention are described in greater detail in the non-limiting Examples that follow.

Example 1

Synthesis and Purification of
2'-Deoxynucleoside 5'- α -Borano-Triphosphates

5 A representative α -P-borane-substituted
2' deoxynucleoside 5'-triphosphate, eg, α -P-borano-
thymidine 5'-triphosphate has been synthesized by Tomasz
et al (*Angew. Chem. Int. Ed. Engl.* 31:1373 (1992)). The
crucial step in the synthesis of the borano-

triphosphates is the attachment of the BH_2 group to the P atom. This reaction was performed at the stage of nucleoside 5'-phosphoramidite by using an exchange reaction with an amine-borane. The N_1O^3' -acylated
5 2'-deoxynucleoside was phosphitylated by (2-cyanoethyl) (N,N-diisopropyl)phosphoramidic chloride to the respective 5'-phosphoramidite derivative. The 5'-phosphoramidite was treated *in situ* with excess pyridine-borane in DMF at 45-50°C for 24 h. (Of the
10 amine-boranes studied, pyridine-borane was the only one that selectively entered into reaction with the phosphorus and did not reduce the base residues.) After ammoniacal treatment, 2'-deoxynucleoside 5'-borano-N,N-diisopropylphosphoramidate was isolated by ion-
15 exchange column chromatography. Reaction of these phosphoramidate derivatives with excess bis-tri-n-butylammonium pyrophosphate gave (after ion-exchange chromatographic purification) the mixture of α -P-borane-substituted 2'-deoxynucleoside
20 5'-triphosphate diastereoisomers in overall yields of 5-6%. The diastereoisomers were separated by HPLC (both diastereoisomers can be incorporated using thermostable polymerases, the stereoisomer that is first eluting on HPLC (Tomasz et al, Angew. Chem. Int. Ed. Engl. 31:1373
25 (1992)), however, appears to be the better substrate for DNA polymerase; both diastereoisomers are resistant to

both 5' and 3' exonuclease, but to different extents depending on the enzyme).

The method of synthesis has been applied to the preparation of α -P borano derivatives of the other three (i.e., A, G, and C) 2'-deoxynucleoside 5'-triphosphates by modification of the procedure.

Example 2

Optimization of Incorporation of dNT^bPs and Exonuclease Digestion

Optimization of reaction conditions (incorporation and digestion) results in a uniform and complete set of bands for each base-specific sequencing lane. Ideally, a single dNT^bP is incorporated into each product molecule such that each position of the PCR product is represented equally. Exonuclease III digestion can be optimized so that each molecule is digested only to, and not beyond, the dNM^bP.

A. Incorporation.

The incorporation efficiency (V_{\max}/K_m) of each of the dNT^bPs can be determined by kinetic analysis (Randall et al, *J. Biol. Chem.* 262:6864 (1987); Boosalis et al, *J. Biol. Chem.* 262:14689 (1987)). The kinetic

parameters have been determined for a different type of boronated dNTP, the base-boronated N'-cyanoborane-2'-deoxyguanosine 5'-triphosphate. For incorporation by exonuclease-free Klenow, it has been found that the
5 base-boronated 2'-deoxyguanosine 5'-triphosphate has a similar V_{max} but a lower K_m than the normal dGTP. For the present invention, a larger or smaller K_m for the dNT^bPs with respect to the normal dNTPs can be compensated for by a change in the ratio of the
10 concentrations of normal to boronated triphosphates in the PCR reaction. As is shown in Example 3, the boronated triphosphates appear to be governed by such a K_m -dominant incorporation, because the appropriate changes in the ratio of [dNTP] to [dNT^bP] during PCR
15 amplification results in uniform and faithful incorporation.

The kinetic parameters for incorporation by Vent or other heat stable DNA polymerases can be determined and the incorporation parameters for all four dNT^bPs thereby
20 rationally optimized. Alternatively, the ratios of boronated to normal triphosphates and their concentrations can be varied in a trial and error fashion.

Kinetic analysis.

The K_m and V_{max} for incorporation of dNTPs and dNT^bPs can be determined by a labeled primer-template/polyacrylamide gel assay (Randall et al, *J. Biol. Chem.* 262:6864 (1987); Boosalis et al, *J. Biol. Chem.* 262:14689 (1987)). The system consists of a radioactively-labeled synthetic primer annealed to a synthetic template that is designed to code for the selected dNTP or dNT^bP at the first position of primer extension. Primer extension is performed for a range of substrate concentrations. The extension products are separated by denaturing PAGE and quantitated on a Molecular Dynamics phosphorimager. At steady state conditions, the initial reaction velocities (v_i) can be determined from the integrated band intensities. The intensity of the extended product (I_1) divided by the intensity of the unextended primer (I_0) plus one half the intensity of the extended product ($0.5 I_1$) gives a measure of v_i (Petruska et al, *Proc. Natl. Acad. Sci. USA* 85:6252 (1988)).

An example of the protocol that can be followed to determine the kinetic parameters for the base-boronated N⁷-cyanoborane 2'-deoxyguanosine 5'-triphosphate (^{7b}dGTP) is as follows. For the normal and boronated dGTP, a primer (5'-CAGGAACAGCTATGGCCTCA-3'; 30 pmol) is end-labeled with 10 μ Ci of γ^{33} P-ATP, annealed to an equal amount of a template (5'-GTGTAGCTGAGGCCATAGCTGTTCTG-3';

30 pmol), and mixed with Vent or Taq DNA polymerase (conditions for use with Taq polymerase are as described below except as otherwise noted in brackets) in buffer A (10 mM Tris-HCl, pH 8.9, 50 mM NaCl, and 0.1% Triton X-100; 32.5 μ l). The primer is extended by mixing 5 μ l of the primer/template duplexes with 5 μ l of various concentrations of dGTP or 3 HdGTP in buffer B (10 mM Tris-HCl, pH 8.9, 50 mM NaCl, 0.1% Triton X-100, and 10 mM MgSO₄) [10 mM Tris-HCl, pH 8.9; 50 mM NaCl and 5 mM MgCl₂]. The reaction is carried out for 1.5 min at 76°C [72°C] and stopped by addition of 10 μ l of loading buffer (95% formamide, 20 mM Na₂EDTA, 0.1% bromophenol blue, 0.1% xylene cyanol). The samples are separated on a 16% polyacrylamide/7 M [8 M] urea sequencing gel and the intensities of the bands are quantitated on a Molecular Dynamics phosphorimager. The K_m and V_{max} can be calculated from the initial reaction velocities by nonlinear regression analysis. To verify that the initial velocities are obtained at an enzyme-limiting condition, the amount of enzyme is adjusted to produce about 20% extension at the highest substrate concentrations.

Time course reactions.

That the initial velocities are obtained during the period of linear accumulation of product can be verified by performing a time course experiment for the highest concentrations of substrate. Labeled primer/template and Vent DNA polymerase in buffer A (32.5 μ l) are mixed with dGTP or 75 dGTP in buffer B (32.5 μ l) at 76°C. Aliquots are withdrawn at 30 sec intervals and mixed with loading buffer. The intensities of the bands can be quantitated and a percent extension vs. time plot can be generated.

Additional considerations.

Vent polymerase includes a 3' exonuclease activity that results in a higher incorporation fidelity relative to exonuclease-free heat-stable polymerases (Eckert and Kunkel, *PCR Methods and Application* 1:17 (1991)). However, due to the additional exonuclease activity, the kinetic analysis measures apparent K_m and V_{max} values that are functions of both the polymerization and the exonuclease activities. The apparent kinetic parameters should aid in optimization of the procedure of the present invention because the relative kinetic parameters need only be compared among the boronated and normal dNTPs (Rappaport, *Biochemistry* 32:3047 (1993);

Singer et al, *Biochemistry* 28:1478 (1989)). It is also noted that the polymerization temperature may cause denaturation of the synthetic primer/template duplex. If it proves to be a problem, the length of the
5 oligomers can be increased.

B. Exonuclease digestion.

Exonuclease digestion can be optimized so that each PCR product is digested only to, and not beyond, the first dNMP. To render the present method a truly one-
10 step procedure, the digestion should take place immediately following amplification and directly in the PCR reaction mixture.

In preliminary experiments, the buffer, time, and temperature of the exonuclease reaction have been
15 adjusted. Exonuclease III from Bethesda Research Laboratories was found to be superior to that from other vendors (Guo and Wu, *Nucl. Acids Res.* 10:2065 (1982); Henekoff, *Gene* 28:351 (1984)). Enzyme concentration, time, and temperature of digestion were determined
20 empirically. The buffer composition was determined by modifying the manufacturer-supplied buffer (50 mM Tris-HCl, pH 8.0, 5 mM MgCl₂, and 1 mM DTT). Tris-HCl concentration and pH were varied systemically and 50 mM and pH 7.5 were found to be optimal. Twenty-five units
25 of exonuclease III in a buffer containing 50 mM Tris-

HCl, pH 7.5, 5 mM DTT, and 5 mM MgCl₂ digested satisfactorily the PCR products in 45 min at 37°C. One hundred units of enzyme effected satisfactory digestion in 30 min under the same conditions, however, in the case of dCM^bP-containing samples, 25 units of enzyme is sufficient. Under these conditions, almost all of the normal dNMPs were digested while the boronated dNMPs were not.

While the preliminary experiments described herein were performed on PCR products that had been ethanol-precipitated prior to exonuclease treatment, conditions can be established that allow direct digestion of the PCR products.

Example 3

Amplification and Sequencing of Region of M13mp2 Phage DNA

Materials:

The 2'-deoxynucleotides dA, dT, dC, and dG 5'- α -borano-thiotriphosphates were synthesized by the methods described in Tomasz et al, *Angew. Chem. Int. Ed. Engl.* 31:1373-1375 (1992)). Oligonucleotides were synthesized on an ABI 380B DNA synthesizer, purified by denaturing PAGE, and recovered by electroelution. The thermostable

DNA polymerase, Vent, was purchased from New England Biolabs. M13mp2 DNA was provided by Ted Gonzalez, Duke University. (Frederico et al, *Biochemistry* 29:2532 (1990)). $\gamma^{33}\text{P}$ -ATP (>1000 Ci/mmol) was purchased from
5 Amersham.

Labeling of Primer 1:

Primer SS20 (5'-TATCGGCCTCAGGAAGATCG-3' complementary to positions 6467 to 6448 of M13mp2; 20 pmol) was 5' end-labeled with $\gamma^{33}\text{P}$ -ATP (20 μCi) and
10 polynucleotide kinase (New England Biolabs; 10 units) in the manufacturer-supplied buffer (10 μl).

Polymerase Chain Reaction:

(+)M13mp2 DNA template (0.5 pmol) was mixed with labeled primer SS20 and unlabeled primer 2 (5'-
15 TCACACAGGAAACACTATGC-3'; positions 6200-6221 of M13mp2; 20 pmol each), dATP, dTTP, dCTP, and dGTP (100 μM of each), and one of each dATP^bP (5 μM), dTT^b1P (5 μM), dCT^bP (100 μM), or dGT^bP (2.5 μM) in PCR buffer (50 mM NaCl, 2 mM MgSO₄, 0.1% Triton X-100, and 10 mM Tris-HCl, pH
20 8.9, at room temperature). The reaction mixture was heated to 95°C for 1 min and returned to ice. Vent DNA polymerase (0.5 μl ; 1 unit) was added and the PCR was performed in an Ericomp thermal cycler for 25 cycles of 95°C for 1 min, 56°C for 1 min, and 76°C for 1 min.

Exonuclease III digestion:

Following PCR amplification, the DNA was ethanol-precipitated, then resuspended in 10 μ l of deionized water. An aliquot (5 μ l) was digested with exonuclease
5 III (25 units) in buffer (50 mM Tris-HCl, pH 7.5, 5 mM DTT, 5 mM MgCl₂; 10 μ l total) for 45 min at 37°C.

Electrophoresis:

Denaturing loading buffer (95% formamide, 0.1% bromophenol blue, 0.1% xylene cyanol, and 20 mM Na₂EDTA;
10 10 μ l) was added to each sample which was then loaded onto a 12% polyacrylamide/8 M urea gel and run for 2 hours at 75 W in TBE buffer (89 mM Tris-borate, 2 mM Na₂EDTA, pH 8.0). The gel was dried under vacuum and the signal was detected by autoradiography.

15 As shown in Figure 2 (see also Figure 3), the results obtained, using conditions that had not been optimized, revealed that the boronated dCT^bP produced a uniform and faithful ladder for 70 bases past the primer, while dG, dA, and dT 5'- α -borano-triphosphates
20 produced a strikingly uniform and faithful sequencing ladder beginning at the 3' end of the labeled SS20 primer and extending about 195 bases into the M13 genome.

The sequencing ladders for dG, dA, and dT 5'- α -borano-triphosphates were clear, uniform, and easy to read. The only ambiguities were:

- 5 dGT^bP - a missing band at position 187 and an
 extra band at position 93.
- dTT^bP - missing bands at positions 92, 126, and
 142 and extra bands at positions 34 and
 35.
- 10 dAT^bP - a missing band at position 65 and extra
 bands at positions 73 and 87-89.

For the 195 bases (215 bases past the 20 base primer) that were sequenced by dG, dT, and dA, each position was scored for each 2'-deoxynucleotide (that is, the presence or absence of a band at the correct positions). Therefore, the accuracy for each base in this preliminary experiment was:

15

- dGT^bP - 99.0% correct.
- dTT^bP - 97.5% correct.
- dAT^bP - 97.5% correct.
- 20 dCT^bP - 100% correct for 70 bases.

Missing bands in the sequencing ladders were probably due to sub-optimal incorporation conditions. By determining the kinetic parameters for incorporation of each dNT^bP, using protocols such as those described in Example 2, optimal conditions for incorporation can be employed in the one-step procedure that should

25

eliminate sequencing errors due to missing bands and allow for longer reads. Alternatively, use of another boronated modified base could be employed.

5 Extra bands in the sequencing ladders were probably due to incomplete exonuclease III digestion. The correct conditions for digestion can be determined by simple time course of digestion experiments, if necessary, the kinetic parameters for exonuclease III digestion can be determined by the labeled primer-
10 template/polyacrylamide gel assay (Otto et al, *Biophys. J. Abstracts*, 37th Annual Meeting, Tu-Pos278, pp. A181). Alternatively, another 3' to 5' exonuclease can be employed.

The incorporation and/or exonuclease III digestion
15 properties of dCT^bP seem to be different from other three dNT^bPs. It is possible that the relative incorporation efficiency of dCT^bP is low, resulting in too few incorporated sequence delimiters. Alternatively, it is possible that, once incorporated,
20 the dCM^bP residues are not as resistant to exonuclease III digestion, thereby allowing digestion of the longer fragments. It is also possible that the dCT^bP preparation was not pure or that the compound could have degraded before use. However, after optimization of
25 both incorporation and exonuclease III digestion, dCT^bP

can be expected to produce longer reads of sequencing data.

Example 4

- 5 (A) Incorporation of Boronated Nucleotides
in PCR Products and (B) Base Specific Resistance
of Extended Primers to Exonuclease Activity

A. Incorporation of Boronated Nucleotides

10 *Materials.* The 5'- α -borano-triphosphates of dA, dT, dC, and dG were synthesized and the diastereomers were HPLC-separated by modifications of the methods described in Tomasz et al (Angew. Chem. Int. Ed. Engl. 31:1373 (1992)). Normal oligonucleotides to be used as primers were purchased from Genset, purified by TLC (USB), and recovered by lyophilization. The
15 thermostable DNA polymerases, Vent and Taq, were purchased from New England Biolabs and Boehringer Mannheim. γ - ^{33}P -ATP and α - ^{33}P -dATP (>1000 Ci/mmol) were purchased from Amersham.

20 *Polymerase Chain Reaction.* Phage T7 DNA template (5 ng) was mixed with 20 pmol primers (T7_{for}: 5'-GGAGCGTAGGAAATAATAC-3' and T7_{rev}: 5'-CGGTTTTTAATTACGTTAGCC-3'), dATP, dTTP, dCTP, and dGTP (100 μM of each), and one of each dATP^b, dTT^b, dCT^b, or dGT^b (2.5 μM each)

in Vent PCR buffer (50 mM NaCl, 2 mM MgSO₄, 0.1% Triton X-100, and 10 mM Tris-HCl, pH 8.9 at room temperature). The reaction mixture was heated to 95°C for 1 min and returned to ice. Vent DNA polymerase (1 µL; 2 units) was added and the PCR was performed in an Ericomp thermal cycler for 25 cycles of 95°C for 1 min, 53°C for 1 min, and 76°C for 1 min. An aliquot of each extended primer-template duplex (10 µL) was digested with exonuclease III (0.5 µL, 32.5 units) for 30 min at 37°C.

Electrophoresis. Samples were separated on a 1% agarose gel.

From Figure 4, it can be seen that both normal and boronated nucleotides produced comparable amounts of DNA (compare lane 1 - all normal, with lanes 3, 5, 7 and 9 - partially boronated). A full length product (509bp) is present in all lanes, indicating that boron does not inhibit PCR amplification.

To demonstrate that the PCR product indeed contained boronated nucleotides and to determine if these nucleotides conferred resistance to nuclease, the PCR product was digested with exonuclease III as seen in Figure 4. Upon digestion, the normal DNA was digested extensively (lane 2), whereas the boron-containing DNA was digested very little, demonstrating that boron was incorporated into the DNA (lanes 4, 6, 8 and 10).

B. Base Specific Resistance of Extended Primers to
Exonuclease Activity

5 *Labeling.* Primer (5'-CAGGAACAGCTATGGCCTC-3';
10 pmol) was 5' end-labeled with γ -³³P-ATP (20 μ Ci) and
polynucleotide kinase (10 units) in the manufacturer-
supplied buffer (10 μ L).

10 *Extension and digestion.* Labeled primer was
annealed to an equal amount of unlabeled template (5'-
GTGTAGCTGAGGCCATAGCTGTTCTG-3'; 10 pmol). Extension was
accomplished with T4 DNA polymerase (1 unit) in the
presence of (a) all normal dNTPs or (b) successive
replacement of one dNTP with the corresponding dNT^bP
(50 μ M each in buffer: 50 mM Tris-HCl, pH 7.5, 10 mM
MgCl₂, 1 mM DTT; 25 μ L) for 10 min at 37°C. An aliquot
15 of each extended primer-template duplex (10 μ L) was then
digested with exonuclease III (0.5 μ L; 32.5 units) for
30 min at 37°C.

20 *Electrophoresis.* Denaturing loading buffer (95%
formamide, 0.1% bromophenol blue, 0.1% xylene cyanol,
and 20 mM Na₂EDTA; 10 μ L) was added to each sample which
was then loaded onto a 16% polyacrylamide/8 M urea gel
and run for 2 hours at 100 W in TBE buffer (89 mM Tris-
borate, 2 mM Na₂EDTA, pH 8.0). The gel was dried under
vacuum and the signal was detected by autoradiography.

25 As seen in Figure 5, the primer was extended in the
presence of all-normal nucleotides and, equally as well,

by successive replacement of each nucleotide by 100% of the corresponding boronated nucleotide. In each case, the primer was extended the full length of the template (lane 2 - all-normal, lanes 4, 6, 8 and 10 - boronated).

5 The presence of a significant quantity of boronated nucleotide (equal here to 4-11% of the total nucleotide concentration in the extended primer) did not affect the electrophoretic mobility of the extended primers.

Following extension, an aliquot of each sample was
10 digested with exonuclease III. The all-normal product was digested to the limit of a duplex substrate (exonuclease III requires double stranded substrate, lane 3), whereas for each case in which a dNT^bP was substituted for a normal dNTP, exonuclease digestion was
15 halted at the position of the boronated substitution (lanes 5, 7, 9 and 11).

The boronated dA, dT, and dG (lanes 5, 7 and 9, respectively) were completely resistant to exonuclease III under these conditions, while the boronated dC was
20 more susceptible to digestion (lane 11). This sensitivity parallels that seen for normal dC (Linxweiller et al, Nucl. Acids Res. 10:4845 (1982)). The differential sensitivity of dC has been compensated for by adding more dCT^bP (50 μ M) to the sequencing
25 reactions and by reducing the amount of exonuclease III from 100 units to 25 units. Thus, the above experiments

demonstrate that (1) the boronated nucleotides allow for primer extension (where 3 out of 8 nucleotides, or 37%, of the newly synthesized product was boronated), (2) the boronated nucleotides are much more resistant to
5 exonuclease III than normal nucleotides, and (3) the base-specific resistance implies that the bases are incorporated correctly.

Example 5

One-Step Sequencing with End-Labeled Primer

10 *Labeling of Primer 1.* Primer T7_{for} (5'-GGAGCGTAGGAAATAATAC-3', positions 34534 to 34552 of phage T7) or primer T7_{rev} (5'-CGGTTTTAATTACGTTAGCC-3'; complementary to positions 35042 to 35025 of phage T7; 20 pmol each) was 5' end-labeled with γ -³³P-ATP (20 μ Ci)
15 and polynucleotide kinase.

Polymerase Chain Reaction. Phage T7 DNA template (5 ng) was mixed with 20 pmol of labeled and unlabeled primers, dATP, dTTP, dCTP, and dGTP (100 μ M each), and one of each dATP^bP (2.5 μ M), dTT^bP (10 μ M), dCT^bP (10 μ M)
20 or dGT^bP (2.5 μ M) in PCR buffer (50 μ L). The reaction mixture was heated to 95°C for 1 min and returned to ice. Vent DNA polymerase (1 μ L; 2 units) was added and the PCR was performed in an Ericomp thermal cycler for

25 cycles of 95°C for 1 min, 53°C for 1 min, and 76°C for 1 min.

Exo III digestion. Following PCR amplification, the DNA was extracted with chloroform:isoamyl alcohol (24:1), precipitated, and resuspended in deionized water (10 µL). An aliquot (5 µL) was digested with *exo III* (25 units) in buffer (50 mM Tris-HCl, pH 7.5, 5 mM DTT, 5 mM MgCl₂; 10 µL total) for 15 min at 37°C.

Electrophoresis. Denaturing loading buffer was added to each sample which was then loaded onto an 8% polyacrylamide/8 M urea gel and run for 2 hours at 100 W in TBE buffer. The gel was dried under vacuum and the signal was detected by autoradiography.

Results in Fig. 6 show that the sequence of about 350 base pairs (second loading not shown) of a 509 bp PCR product could be read. The α-P-borano dNTPs are thus incorporated faithfully into the correct positions of the DNA and, once incorporated, code for the correct complementary 2'-deoxynucleotide. If this were not true, then the background would have been blurry and the correct sequence could not have been obtained. The α-P-borano 2'-deoxynucleotides thus render the PCR products resistant, base-specifically, to exonuclease III, as evidenced by the correct sequencing ladder. By incorporating 2'-deoxynucleoside 5'-α-P-borano-triphosphates into base-specific PCR reactions and then

digesting the PCR products with exonuclease III, a uniform series of bands was produced, thereby defining the sequence of approximately 350 bases of a 509 base-pair PCR product.

5 The sequence data were remarkably clear and the bands were of quite uniform intensity. However, as is shown in Figure 6A, there were two troublesome regions; one near position 34600 of the forward sequence and one near position 34915 of the reverse sequence, with bands
10 across all four lanes. Because there were no pauses in the polymerization reaction (data not shown), it was hypothesized that these extra bands were most likely due to incomplete digestion. When the experiment was repeated, using samples that were digested with 130
15 instead of 25 units of exonuclease III (Figure 6B), the extra bands near position 34600 of the forward strand disappeared completely, producing an easily readable sequence. The region surrounding position 34915 of the reverse strand was also clearer than had been observed
20 previously; however, the correct sequence could not be read without ambiguity. Likely, there is a secondary structure associated with this region because (1) reagents, such as DMSO, which disrupt secondary structure also help to alleviate the sequence
25 ambiguities, (2) with normal nucleotides, the exonuclease III pauses at this site, and (3) the pause

is specific to this region and not to other sequences examined to date. The nuclease resistance is strand-specific, however, and thus the sequence of the other strand can be read. One alternative approach is to
5 raise the temperature of the digestion, and thus melt out the secondary structure. If the exonuclease digestion were to be performed at high temperature, the secondary structure would be expected to be disrupted and digestion would produce uniform fragments at all
10 sites.

Example 6

Bi-directional One-Step PCR Sequencing with a Biotinylated Primer

Polymerase Chain Reaction. Phage T7 DNA template
15 (10 ng) was mixed with 20 pmol of biotinylated (ϕ 9_{for} 5'-ACAGCTTCACCTGAGGCTATGG-3'; positions 21786-21807) and unmodified (ϕ 9_{rev} 5'-TCTTCGTTCTCCTCGTACTCACGC-3'; complementary to positions 22391-22414; 629 bp product) primers, dATP, dTTP, dCTP, and dGTP (100 μ M of each),
20 one of each dAT^bP (2.5 μ M), dTT^bP (10 μ M), dCT^bP (10 μ M), or dGT^bP (2.5 μ M), and 13 μ Ci α -³³P-dATP in Taq PCR buffer (10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1.5 mM MgCl₂; 50 μ L). The reaction mixture was heated to 95°C for

1 min and returned to ice. *Taq* (5 units) and Deep Vent (0.005 units) DNA polymerases were added and the PCR was performed in an Ericomp thermal cycler for 25 cycles of 95°C for 15 sc, 60°C for 1 min, and 76°C for 1 min.

5 *Binding to magnetic beads.* An aliquot of the PCR reaction (20 μ L) was mixed with streptavidin-linked magnetic beads (Dynal; 10 mg/mL in 5 mM Tris-HCl, pH 7.5, 0.5 mM EDTA, 1 M NaCl; 20 μ L) and incubated with mild agitation for 45 min.

10 *Exonuclease III digestion.* The PCR products bound to the magnetic beads were immobilized by a magnet and washed twice with exonuclease III buffer. The samples were resuspended in exonuclease buffer (10 μ L) and digested with exonuclease III (60 units) for 30 min at
15 37°C.

20 *Isolate unmodified primer strand fragments.* After digestion, the samples were heated for 1 min at 95°C to denature the PCR products. The magnetic beads, along with the biotinylated primer strand fragments, were immobilized with a magnet and the unmodified primer strand fragments were removed in the exonuclease buffer. The fragments were mixed with an equal volume of loading buffer and loaded buffer and loaded (6 μ L) onto a sequencing gel (8% polyacrylamide/8 M urea).

25 *Isolate biotinylated primer strand fragments.* The biotinylated primer strand fragments were resuspended in

loading buffer (10 μ L) and heated for 1 min at 95°C to detach them from the streptavidin-linked magnetic beads and then loaded (3 μ L) onto a sequencing gel.

Electrophoresis. Performed as described for Figure 6.

Cycle sequencing. As a control, cycle sequencing was performed on T7 DNA as directed in the Perkin Elmer Amplitaq cycle sequencing kit. The primers and T7 template DNA was the same as described below for the one-step method except that 1000 ng of highly purified template DNA was used for each cycle sequencing sample whereas only 10 ng were required for the present method.

A region of T7 DNA extending from the phi 9 promoter (629 bp product) was sequenced bi-directionally using the biotinylated primer - magnetic beads strategy (see Figure 7). For comparison, the same region was sequenced using labeled primers by conventional cycle sequencing (see Figure 7). For each method, the sequence could be read from approximately A₂₁₉₀₀ to G₂₂₃₅₀, or about 200-300 bases per strand from a single loading. No secondary structure-induced artifacts were noted for either the one-step or cycle sequencing method; however, each method had at least one position where the correct base could not be called because of an extremely weak band. In each case where a particular base could not be determined by one method, the base could be called

correctly by the other method. The desirability of determining a sequence by two independent methods has been noted previously (Koop et al, Biotechniques 14:442 (1992)).

5

Example 7

Suitability of Boronated Deoxynucleotide for Direct Gene Transfer

Figure 8 shows that α -borano triphosphates can be incorporated by PCR into a duplex which contains a promoter (T7 phi 17) and a short downstream flanking region. When the boronated templates were used in an *in vitro* transcription system, the boronated templates were shown to produce RNA when the templates contained up to 75% boronated nucleotides. From Figure 8 Upper panel, the correct 110 base PCR product was produced using, along with the other normal dNTPs, boronated dGTP (lane 3), boronated dCTP (lane 4), boronated dA (lane 5), boronated dG+dC (lane 6), boronated dC+dA (lane 8), and boronated dG+dC+dA (lane 9). Each of the PCR templates was capable of being transcribed into RNA and the amount of RNA was roughly proportional to the amount of DNA produced by PCR (corresponding lanes of the lower panel). Thus it appears that boronated deoxynucleotides

do not inhibit transcription and therefore are suitable for use in direct gene transfer.

Example 8

Resistance of Boronated DNA to Serum and Exonuclease
5 Digestion, Transcription of Boronated DNA into mRNA
and Translation of mRNA into Protein

A. Resistance to degradation: Equal amounts of normal and boronated (prepared using 100% boronated dCTP) PCR products (full length luciferase gene PCR amplified from
10 plasmid (PROMEGA L482B); lucfor
5'TTGATGGAGCTGATAACCGCTCG3', lucrev
5'ATCTTCCCCATCGGTGATGTCG3'; Taq buffer: 10mM Tris (8.3),
50mM KCl, 2.5mM MgCl₂; dNTPs: 100 micomolar each, Taq 5
units; plasmid DNA 5ng; 25 cycles: 95 degrees C (15
15 min.), 60 degrees C (45 min), 72 degrees C (1 min) and
repeat) were incubated with 1/10 volume of mouse serum
in buffer (50 mM NaCl, 5 mM MgCl₂, 5 mM DTT and 10 mM
Tris, pH 7.6) at 37°C for 20 min or for 40 min. As
shown in Figure 9A, full-length normal DNA was digested
20 completely by 20 minutes (compare lane 2 (20 min) and
lane 3 (40 min) with lane 1 (untreated)). Full length
boronated DNA, however, persisted throughout the 40 min

incubation (compare lane 5 (20 min) and lane 6 (40 min) with lane 4 (untreated)).

Approximately equal amounts of normal and boronated (prepared using 100% boronated dG or dC) PCR products (as above) were labeled and separated on a denaturing polyacrylamide gel (see Fig. 9B (lane 1 normal, lane 2 100% boronated dG, lane 3 100% boronated dC). Aliquots of each PCR product were digested with exonuclease III. As shown in Fig. 9B, normal DNA was digested by exonuclease III (compare lane 4 with lane 1), while boronated DNA was resistant to digestion (compare lanes 5 (dG^b) and 6 (dC^b) with lanes 2 and 3, respectively).

B. Transcription of boronated DNA into mRNA:

Aliquots of PCR product (as above) were transcribed into mRNA using $\alpha^{35}\text{S}$ -ATP and the Promega TNT transcription/translation system. As shown in Fig. 9B, normal (lane 7) and boronated (lane 8 - (100%)dG^b; lane 9 (100%) dC^b) were transcribed into full-length mRNA.

C. Translation of mRNA transcript into protein:

Aliquots of normal and boronated PCR products (as above) were translated into protein using ^{35}S -methionine and the Promega TNT transcription/translation system. The labeled protein was separated on an SDS-

polyarylamide gel. As shown in Fig. 9C, full length protein was produced by both the normal (lane 1) and boronated (lane 2 - boronated (100%) dG; lane 3 - boronated (100%) dC) template.

5 In a separate study, normal and boronated templates were shown to be translated into functional enzyme at the same rate. PCR products containing all-normal, 100% boronated dG, or 100% boronated dC were transcribed/translated into functional luciferase enzyme
10 using the Promega TNT system. Aliquots were withdrawn from the reaction mixture at five minute intervals and frozen on dry ice. The samples were thawed on wet ice and aliquots (0.5 μ L) were mixed with luciferin assay reagent (10 μ L; Promega). Light emission from the assay
15 was quantified in a scintillation counter. Light emission ($\sqrt{\text{cpm} - \text{background}}$) was converted to luciferase concentration by comparison to a standard curve and plotted vs. time. As shown in Fig. 10, active luciferase was produced by normal and boronated
20 templates at the same rate.

Example 9Resistance of Boronated DNA to
Digestion by 5' Exonuclease

Normal and boronated PCR products were prepared as
5 described in Example 6 (Taq only; primers: phi9for,
phi9rev (see Example 6); Taq buffer; dNTPs: 100
micromolar, Taq: 5 units; T7 DNA: 5 ng; 25 cycles: 95
degrees C (15min), 60 degrees C (1 min), 72 degrees C (1
min), and repeat) except that, for base-specific
10 boronated samples, dNTP's of the second-eluting (HPLC)
diastereoisomer (50 μ M) were added to the reaction.
After amplification, the samples were supplemented with
MgCl₂ (2.5 mM) and DTT (1 mM). T7 gene 6 (25 units), a
5' exonuclease that requires a double stranded
15 substrate, was added to an aliquot of the PCR reaction
(10 μ L) and allowed to react for 10 min. The samples
were separated on a 1% agarose gel.

As shown in Figure 11, normal DNA was digested to
completion (compare lane 2 with lane 1). In contrast,
20 the boronated samples were resistant to digestion
(compare lanes 4, 6, 8 and 10 with lanes 3, 5, 7 and 9,
respectively).

Example 10

Chemical Synthesis of Oligodeoxynucleotides
Containing Boranophosphate Diester Linkages

Synthesis of dithymidine boranophosphate (4) in
5 solution:

11.5 mg (41 μmol) of 3'-O-acetyl-2'-deoxythymidine
and 38.3 mg (54 μmol , 1.3 eq) of 5'-O-dimethoxytrityl-
2'-deoxythymidine-3'-H-phosphonate were dissolved in
0.5 mL of anhydrous pyridine and 33.2 μL (270 μmol ,
10 6.5 eq) of pivaloyl chloride (PivCl) was added. After
5 min of incubation, 5 mL of CH_2Cl_2 was added and the
reaction mixture was extracted two times with 10 mL of
1 M aqueous KH_2PO_4 . The organic layer was dried over
 Na_2SO_4 , concentrated, and 5'-O-DMT-thymidylyl[3'-5'] (3'-
15 acetylthymidylyl) H-phosphonate (2) was purified by
flash chromatography and precipitated in hexane. (See
Figure 12)

32.0 mg (36.5 μmol) of 5'-O-DMT-thymidylyl[3'-
5'] (3'-acetylthymidylyl) H-phosphonate (2) was dissolved
20 in 0.5 mL of anhydrous 1,4-dioxane and 27 μL (110 μmol ,
3 eq) of bis(trimethylsilyl)acetamide was added. After
6 hours of incubation at room temperature (when ^{31}P NMR
spectroscopy showed complete conversion to phosphite
triester), 70 μL (560 μmol , 15 eq) of 8 M pyridine-

borane complex (Py:BH₃) in pyridine was added. The boronation reaction was allowed to proceed 16 hours at 50°C. ³¹P NMR spectroscopy showed complete disappearance of phosphite triester and formation of boranophosphate triester (3). Then 0.5 mL of concentrated ammonia was added to the reaction mixture. After 2 hours of incubation at room temperature, the desired product (4) was isolated by reverse-phase HPLC. Yield: based on ³¹P NMR spectroscopy - 60%, on HPLC trace - 50%.

10 *Synthesis of dimethyl boranophosphate in solution:*

4.6 µL (5.5 mg, 50 µmol) dimethyl H-phosphonate was dissolved in 1 mL of anhydrous 1,4-dioxane and 124 µL (500 µmol, 10 eq) of bis(trimethylsilyl)acetamide was added. After 3.5 hours of incubation at room temperature (³¹P NMR spectroscopy showed complete conversion to phosphite triester), 125 µL (1 mmol, 20 eq) of 8 M pyridine-borane complex in pyridine was added. The boronation reaction was allowed to proceed 16 hours at 50°C. ³¹P NMR spectroscopy showed complete disappearance of phosphite triester and formation of boranophosphate triester. Then 1.0 mL of concentrated ammonia was added to the reaction mixture. After 2 hours of incubation at room temperature, a new signal (quartet centered at 92.3 ppm) appeared corresponding to

the desired dimethylboranophosphate. Yield based on ^{31}P NMR spectroscopy was 85%.

Manual synthesis of dithymidine boranosphosphate (4) on solid support:

5 100 mg (130 μmol of nucleoside/g) 5'-
dimethoxytrityl-2'-deoxythymidine-3'-succinyl silica gel
support or 50 mg (280 μmol of nucleoside/g) 5'-
dimethoxytrityl-2'-deoxythymidine-3'-succinyl
polystyrene support was treated with 300 μL of 2.5%
10 dichloroacetic acid in dichloromethane for 1.5 min,
washed with 300 μL of acetonitrile 2 times and 300 μL
of acetonitrile:pyridine (1:1) mixture. Then 400 μL of
0.1 M 5'-O-dimethoxytrityl-2'-deoxythymidine-3'-H-
phosphonate (40 μmol , 3 eq) in acetonitrile:pyridine
15 (1:1) mixture and 25 μL (200 μmol , 15 eq) of pivaloyl
chloride were added. After 3 min of coupling, the
support was washed with 300 μL of acetonitrile 2 times,
treated with 300 μL of 2.5% dichloroacetic acid in
dichloromethane for 1.5 min and washed again with 300 μL
20 of acetonitrile 2 times. Then 33.5 μL (140 μmol , 10 eq)
of bis(trimethylsilyl)acetamide in 0.5 mL of anhydrous
1,4-dioxane was added. After 6 hours of incubation at
room temperature, 35 μL (280 μmol , 20 eq) of 8M
pyridine-borane complex in pyridine was added and the
25 boronation reaction was allowed to proceed 16 hours at

50°C. After the boronation was complete, the support was washed with 400 μ L of 1,4-dioxane, 1 mL of water and the product was cleaved from the resin and deprotected by treatment with concentrated ammonia during 2 hours at room temperature. Dithymidine boranophosphate was purified on RP HPLC. Yield based on chromatography trace was 75% for silica gel support and 60% for polystyrene resin.

Example 11

10 Resistance of Boranephosphonate Oligonucleotides to Restriction Enzymes

Boranephosphonate oligonucleotides are a new class of DNA mimics that show excellent resistance towards exonucleases (Li et al, NAR (1996)). The nonbridging P-BH₃⁻ group of the boranephosphonate backbone is structurally similar to both the P-O⁻ group of normal phosphodiester backbones and P-S⁻ group of phosphorothioate backbones. The relative stability of recognition sequences containing boron-modified phosphodiester backbones towards restriction enzymes have been examined and their stability with P-O and P-S oligonucleotides of the same sequences compared. A set of 57-59 base pair oligonucleotide hybrid duplexes

containing a 44-nucleotide stretch of either
all-boranephosphonate (P-BH₃), all-phosphorothioate
(P-S), or all-normal phosphodiester (P-O) linkages in
one strand and all-normal (P-O) phosphodiester linkages
5 in the opposite strand was synthesized by primer
extension method using Klenow polymerase and
deoxynucleoside- α -P-boranotriphosphate (dNTP- α -BH₃).
The results indicate that the duplex containing
boranephosphonate linkages is more resistant to
10 hydrolysis by EcoRI, EcoRV, and HindIII enzymes than
phosphorothioate oligonucleotides, which in turn are
more resistant to hydrolysis than normal
oligonucleotides. EcoRI and HindIII enzyme recognition
sites containing boranephosphonate linkages were
15 hydrolyzed at least three-fold and five-fold slower than
those of phosphorothioate linkages or phosphate
linkages, respectively. An EcoRV recognition site was
hydrolyzed eight-fold slower than a recognition site
containing phosphorothioate linkages. In contrast, when
20 the boranephosphonate or phosphorothioate linkages were
present only as part of a TaqI recognition sequence, but
not at the scissile phosphodiester bond, there was no
interference with TaqI restriction.

In the present study, a hybrid duplex has been
25 constructed containing a stretch of boranephosphonates
or phosphorothioates on one strand and normal phosphates

on the opposite strand. For comparison, a similar duplex has been created containing normal phosphodiester on both strands. The sequence of the DNA duplex is such that it contains restriction enzyme recognition sites for EcoRI, EcoRV, HindIII, and TaqI enzymes (Figure 13). The lability of boranephosphonates to undergo sequence-specific hydrolysis of restriction enzymes has been investigated and compared with substrates containing normal phosphodiester and phosphorothioates linkages. In general, the boranephosphonate linkages are several-fold more nuclease resistant than phosphorothioates and normal phosphodiester.

Methodology

15 *Oligonucleotide synthesis and end labeling*

The phosphodiester and phosphorothioate oligonucleotides used in this study were synthesized in 0.2 μ mole quantities by the Duke University DNA Synthesis Facility. The crude oligonucleotides were purified by electrophoresis in a 0.4 mm thick 10% polyacrylamide-7M urea gel using 1 X TBE buffer (50 mM Tris-borate pH 8.3, 1mM EDTA). The DNA in the gel was visualized by UV shadowing. The desired DNA band from the gel was isolated by the crush and soak method

(Sambrook et al, Molecular Cloning - A Laboratory Manual Cold Spring Harbor Press, NY (1989)). The concentration of the DNA was determined by measuring the UV absorbance at 260 nm. The amount of DNA recovered by this method was ~30%.

The sequence of the 15-mer primer and the 57-mer template are shown in Figure 13. The purified primer (26.4 pmoles, 1.32 μ M) was 5'-labeled with [33 P- γ]-ATP (28 pmoles, 1.4 μ M, 1060 Ci/mmol, Amersham) and T4-polynucleotide kinase (10 units, NEBiolabs, MA) in 20 μ l at 37°C for 1 hour. After labeling, the labeled oligonucleotides (1.32 μ M, 20 μ l) were separated from unreacted ATP by passing through a NENSORB column (Dupont-NEN products, MA) as per the manufacturers procedure.

Enzymatic synthesis of boranophosphate or phosphorothioate containing oligonucleotides

The primer extension reactions were performed using $[\alpha\text{-BH}_3]\text{-dNTPs}$ or $[\alpha\text{-S}]\text{-dNTPs}$ or normal dNTPs of guanines, cytosines, adenines and thymines. The $[\alpha\text{-BH}_3]\text{-dNTPs}$ were prepared as per the published procedures (Tomasz et al, Angew. Chem. Int. Edn Engl. 31:1373 (1992)). Substitution of one of the non-bridging oxygens by a BH_3 group in the phosphate moiety gives rise to R_p and S_p diastereomers. These

diastereomers of [α]-borano-deoxynucleotide trisphosphates were separated by HPLC (Huang, Ph.D. thesis, Duke University (1994); Li et al, Nucleic Acids Res. 23:4495 (1995)). The faster moving peak (isomer I, putative Rp) of the two isomers was used in this study for primer extension reactions (Li et al, Nucleic Acids Res. 23:4495 (1995)). In the same way, diastereomers of [α -S]-dNTP were separated by HPLC (Grady, Masters Thesis, Duke University (1995)) and the Sp isomer was used to synthesize all stereo regular phosphorothioate oligonucleotides.

Briefly, the primer extension reactions were done as described below. The 5'-end labeled primer (15 nt in length, 4.6 pmoles) was hybridized to a template (57 nt in length, 5.4 pmoles) in 277 μ l buffer containing 52 mM Tris HCl, pH 7.5, 7.5 mM MgCl₂, and 3.7 mM DTT. This mixture was heated to 95°C for 5 minutes and then slowly cooled to 25°C. To this mixture, a solution of the appropriate deoxynucleotide triphosphate mixture (27 μ l, 20 μ M stock, 1.8 μ M final) and 60 units of 3'-5' exonuclease free Klenow polymerase (6 μ l, US Biochemical Corp., Ohio) were added. The mixture was incubated at 37°C for 6 hours. The reaction was stopped with 5 μ l of EDTA (0.5M). To this mixture, 3 M sodium acetate, pH 5.2, was added such that the final concentration of sodium acetate was 0.35 M. The mixture was precipitated

with 3 volumes of ethanol by centrifugation at 15,000g for 30 minutes at room temperature. The precipitate obtained after centrifugation was washed with 700 μ l of ice cold 80% ethanol. The pellet obtained after washing
5 was dried and dissolved in 50 μ l of 90% formamide dye containing xylene cyanol and bromophenol blue dye. The full length primer extension product present in the sample was separated from proteins and incomplete extension product in a 10% polyacrylamide-7M urea gel
10 (0.4 mm x 30 cm x 45 cm) using 1X TBE buffer at 2000 V for 1 hour. The primer extension products were identified by autoradiography. The gel slice (7 x 1.5 cm x 0.4 thick) corresponding to the selected DNA band in autoradiogram was cut from the gel and
15 placed in a sealed pipette tip containing glass wool. The gel slice was crushed and eluted with 1 mL of buffer containing 10 mM Tris HCl, pH 8.0, 100 mM NaCl and 1 mM EDTA. The eluents were concentrated in a Centricon 10 unit (Amicon, Inc., Beverly, MA). This concentrated
20 sample was used directly for further biochemical analysis.

*Restriction analysis of oligonucleotide duplex
containing P-O, P-S, and P-B linkages with HindIII,
EcoRI, EcoRV, TaqI and PvuI enzymes*

Approximately 500-2000 cpm of the gel-purified
5 59mer boranephosphate-modified, phosphorothioate-
modified and control normal duplexes (7 μ l, 5-10 nM)
were incubated with 1.5 to 20 units of EcoRI, EcoRV,
HindIII, PvuI, or TaqI restriction enzyme in a final
volume of 10 μ l digestion buffer. The EcoRI and PvuI
10 digestion was carried out in 100 mM NaCl, 50 mM Tris-HCl
(pH 7.9), 10 mM MgCl₂, 1 mM DTT and 100 μ g/mL BSA at
37°C for 1 hour. The EcoRV restriction was done by
incubating the sample (7 μ l, 5-10 nM) in a buffer
containing 100 mM NaCl, 10 mM Tris-HCl (pH 7.5), 7 mM
15 MgCl₂, 7 mM 2-mercaptoethanol and 100 μ g/mL BSA at 37°C
for 1 hour. The HindIII restriction was carried out in
a buffer containing 50 mM NaCl, 10 mM Tris-HCl (pH 7.9),
10 mM MgCl₂, 1 mM DTT and 100 μ g/mL BSA at 37°C for
1 hour and the TaqI restriction was done in 100 mM NaCl,
20 10 mM Tris-HCl (pH 8.4), 10 mM MgCl₂, 1 mM DTT and 100
 μ g/mL BSA 55°C for 1 hour. The EcoRV enzyme was
obtained from U.S. Biochemicals Corporation, Ohio and
the EcoRI, HindIII, TaqI, and PvuI enzymes were obtained
from NEBiolabs, MA. The reactions were stopped by
25 adding 0.5 M EDTA to a final concentration of 20 mM.
The samples were analyzed in a 10% denaturing

polyacrylamide gel and the amount of radioactivity corresponding to each band in the gel was quantitated using a Molecular Dynamics phosphorimager (model no. 445 SI, Mountainview, CA).

5 Results

Enzymatic synthesis of 59mer duplexes containing 44-nt stretch of normal, phosphorothioate or boranophosphate linkages (P-O, P-S, or P-B duplex)

10 The sequence of the oligonucleotide primer and template are shown in Figure 13. The sequence of the 15-nt long primer is underlined and the bottom strand (template strand) contains 57 nucleotides. The newly incorporated normal or modified nucleotides are shown in italics. In the template/primer duplex, the primer
15 strand has a two-nucleotide overhang at the 5' end to facilitate the differential 3' end labeling of extended primer and template strands. As a result, the length of the primer extended product would be 59 nt long.

20 The restriction enzyme recognition sites present in this sequence include restriction enzyme recognition sites for BamHI, EcoRI, EcoRV, HindIII, PvuI and TaqI enzymes. The PvuI recognition site (5' CGATCG) is present at the 3' end of the primer, which has phosphodiester linkages on both strands. This site

serves as an internal control for the verification of the duplex nature of the primer extension product.

Under the conditions described above, the complete extension of 4.8 pmoles of primer required as low as 5 600 pmoles of modified or normal dNTPs. After primer extension, the full length product was separated from unreacted dNTPs and proteins by denaturing acrylamide gel. The formation of correct-size product was inferred from its electrophoretic mobility in a denaturing gel. 10 The presence of two major bands (migrating with RF of 58 and 59 nt) and one minor band (RF of 57 nt) in all the three lanes (P-O, P-S, and P-BH₃) suggested not quite complete extension of the primer extension reaction.

15 *Restriction of P-O, P-S, and P-B oligonucleotide duplexes with EcoRI, EcoRV, HindIII, PvuI and TaqI enzymes*

EcoRI. The EcoRI digestion of P-O, P-S and P-B DNA duplex with 5' label on the primer strand should result 20 in a 38 nucleotide-long fragment. In fact, a correct-size restriction fragment was observed in the gel due to EcoRI restriction at a single site for the P-O, P-S, and P-B duplexes. Treatment of the boron modified duplex with 20 units of EcoRI showed only ~21% cutting, while 25 the normal phosphate and phosphorothioate containing

oligonucleotides duplexes were restricted by more than 80%. Restriction at a single unique EcoRI recognition site on the boronated strand by EcoRI enzyme also confirms the presence of correct recognition sequence.

5 This result also confirms previous results that boronated deoxynucleoside monophosphates are faithfully incorporated by Klenow DNA polymerase during primer extension reaction. Restriction of a mixture containing equal amounts of boron modified duplex and normal DNA
10 duplex with 1.5 U of EcoRI resulted in a 2-fold reduction in the restriction of normal DNA duplex compared to the restriction of a P-O duplex alone in the mixture. This indicated that the boronated DNA competes
15 the observed slow rate of hydrolysis may not be due to weak binding at the recognition site but due to slow rate of hydrolysis.

HindIII. Similarly, the treatment of boron modified DNA with HindIII enzyme gave little or no
20 restricted product on the modified strand, yet under these conditions there was cleavage at the HindIII site in the P-S and P-O duplexes (Figure 3A, lane 8 and 9, respectively). The restriction in these duplexes at a single HindIII site was confirmed by comparing the
25 mobility of the HindIII restricted DNA sample (17 nucleotide) in a sequencing gel. The amounts of

cutting at the normal phosphodiester linkage or at the phosphorothioate sites by HindIII enzyme were ~80% and ~60%, respectively.

5 TaqI. The DNA duplex tested contains two TaqI restriction sites. (a) For the restriction site located at the juncture of the primer and extended primer, a section of the recognition sequence contains a normal phosphodiester linkage and the other section contains P-S or P-B modified linkages. TaqI restricts all three
10 (P-O, P-S, and P-B) oligonucleotide duplexes at both sites. The TaqI restriction in the primer sequence section and at the modified sequence were confirmed by analysis of the mobility of the products in a sequencing gel and they corresponded to 14 nt and 36 nt long
15 fragments. Both phosphorothioate and boranephosphonate modification did not interfere with the TaqI restriction in this sequence. The TaqI restricts completely at the site present in the primer sequence. (b) The restriction efficiency at the second site could not be
20 determined due to further TaqI restriction on the 36 nt long restricted product results in 14 nt long fragment.

 PvuI. As a further control, it was determined whether cleavage would occur for all three duplexes at a PvuI recognition site which is located completely within
25 the primer part of the DNA duplex. Restriction of the 59mer gel-purified product with PvuI enzyme indicated

complete hydrolysis of the normal, phosphorothioate modified and boronated duplexes. The mobility of the PvuI restriction fragment corresponded to the expected 11 nt long fragment in the DNA size ladder. From this experiment, it was inferred that gel-purified DNA did not contain any extraneous impurities inhibitory to restriction enzymes. The complete restriction of the P-B oligonucleotide duplex by PvuI enzyme further indicated that the boranophosphate linkages which lie outside the PvuI recognition site did not interfere with the enzyme restriction.

The EcoRV restriction of a normal or P-S or P-B hybrid DNA duplex containing 5' end label on the primer strand should result in 34-nt long fragment, since it contains only one EcoRV site. The size of the product obtained after EcoRV restriction was confirmed by comparing the mobility of the product with the mobility of a DNA size ladder. Treatment of the boronated oligonucleotide duplex with 1.5 units to 12 units of EcoRV did not result in any fragment due to EcoRV restriction, whereas treatment of the duplex DNA containing phosphodiester linkage or phosphorothioate linkage with only 1.5 units of EcoRV enzyme gave ~72% cutting and ~40% cutting, respectively.

Restriction of a mixture containing an equal amount of boron modified duplex and normal DNA duplex with

1.5 U of EcoRV resulted in a 3-fold reduction in the restriction of the normal DNA duplex. This reduction in the restriction of a normal DNA duplex with EcoRV enzyme indicated that the boronated DNA duplex may be competing with normal DNA duplex for binding at the cognate site and that the observed nuclease stability of boranephosphonate oligonucleotide with EcoRV enzyme is not due to lack of enzyme binding at the cognate site but due to weak hydrolysis at the recognition site.

10

* *

Documents referenced hereinabove are hereby incorporated in their entirety by reference.

One skilled in the art will appreciate from a reading of the foregoing disclosure that various changes in form and detail can be made without departing from the true scope of the invention.

15

WHAT IS CLAIMED IS:

1. A method of synthesizing an oligonucleotide comprising an internucleotide boranophosphate linkage comprising:

i) synthesizing an oligonucleotide comprising an internucleotide H-phosphonate diester linkage,

ii) converting said H-phosphonate diester linkage to a phosphite triester linkage,

iii) boronating said phosphite triester linkage so that a boranophosphate triester linkage is formed; and

iv) converting said boranophosphate triester linkage to a boranophosphate diester linkage so that said oligonucleotide comprising an internucleotide boranophosphate linkage is formed.

2. The method according to claim 1 wherein said conversion of step (ii) is effected by silylation or halogenation.

3. The method according to claim 1 wherein said conversion step (ii) is effected with bis(trimethylsilyl)acetamide.

4. The method according to claim 1 wherein said conversion of step (ii) is effected under anhydrous and anoxic conditions.

5. The method according to claim 1 wherein said boronation of step (iii) is effected using a boron complex.

6. The method according to claim 1 wherein said boronation of step (iii) is effected using an amine borane.

7. The method according to claim 1 wherein said boronation of step (iii) is effected using a pyridine borane complex.

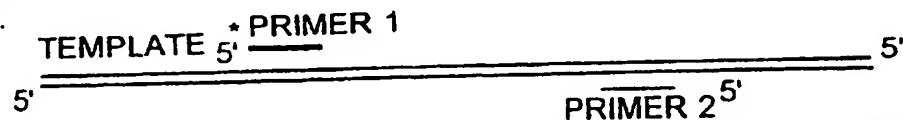
8. The method according to claim 1 wherein said conversion of step (iv) is effected using alkaline conditions.

9. The method accordance to claim 1 wherein said conversion step (iv) is effected using ammonia.

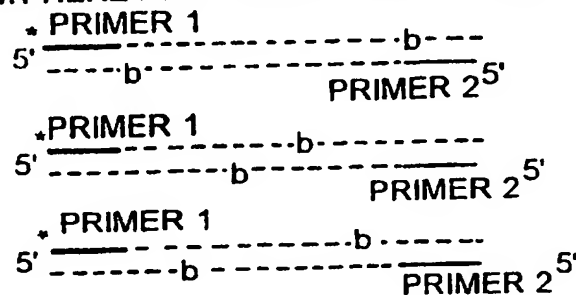
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FIG. 1A**ONE-STEP PCR SEQUENCING
WITH α -BORONATED dNTP'S**

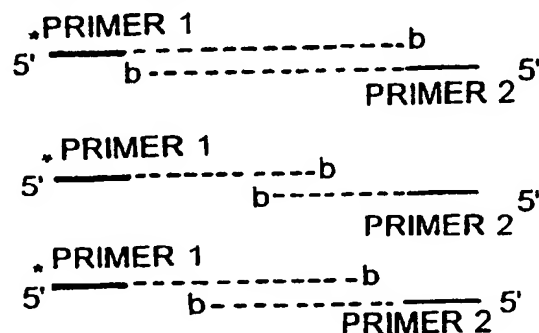
1. ANNEAL LABELED PRIMER 1 AND UNLABELED PRIMER 2 TO DNA TEMPLATE.



2. IN SEPARATE A-, T-, G-, AND C-SPECIFIC REACTIONS, INCORPORATE THE APPROPRIATE BORONATED 2'-DEOXYNUCLEOTIDE BY PCR AMPLIFICATION. SHOWN HERE ARE EXAMPLES OF THREE SUCH PRODUCTS.



3. DIGEST UP TO THE 3' BORONATED dNMP WITH EXONUCLEASE.



4. SEPARATE FRAGMENTS BY PAGE AND READ THE DNA SEQUENCE.

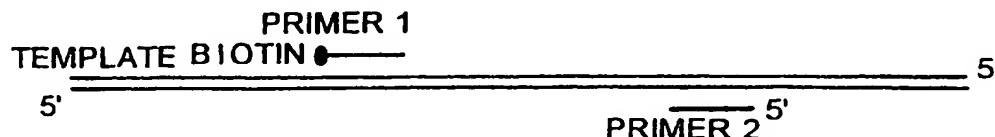
A	T	C	G
—		—	—
—	—	—	—
—		—	—
	—	—	

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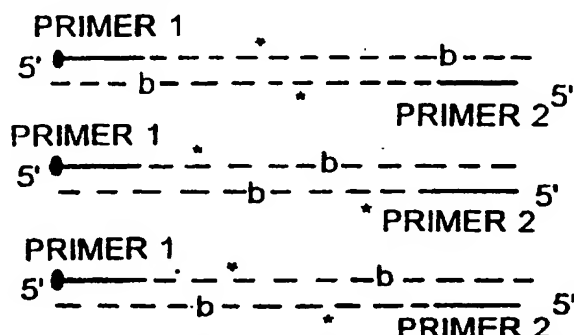
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FIG. 1B**BIDIRECTIONAL ONE-STEP SEQUENCING WITH MAGNETIC BEADS**

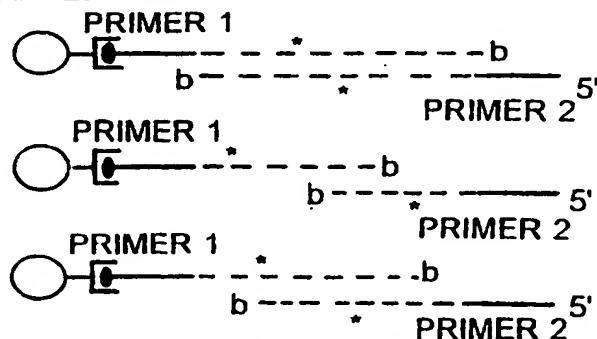
1. ANNEAL BIOTINYLATED PRIMER 1 AND UNMODIFIED PRIMER 2 TO DNA TEMPLATE.



2. IN SEPARATE A-, T-, G-, AND C-SPECIFIC REACTIONS, INCORPORATE BOTH THE APPROPRIATE BORONATED 2'-DEOXYNUCLEOTIDE AND LABEL BY PCR AMPLIFICATION.



3. BIND BIOTINYLATED PCR PRODUCTS TO STREPTAVIDIN-LINKED MAGNETIC BEADS, IMMOBILIZE BEADS TO MAGNET, WASH WITH BUFFER, AND DIGEST BACK TO THE 3' BORONATED dNMP WITH EXO III.



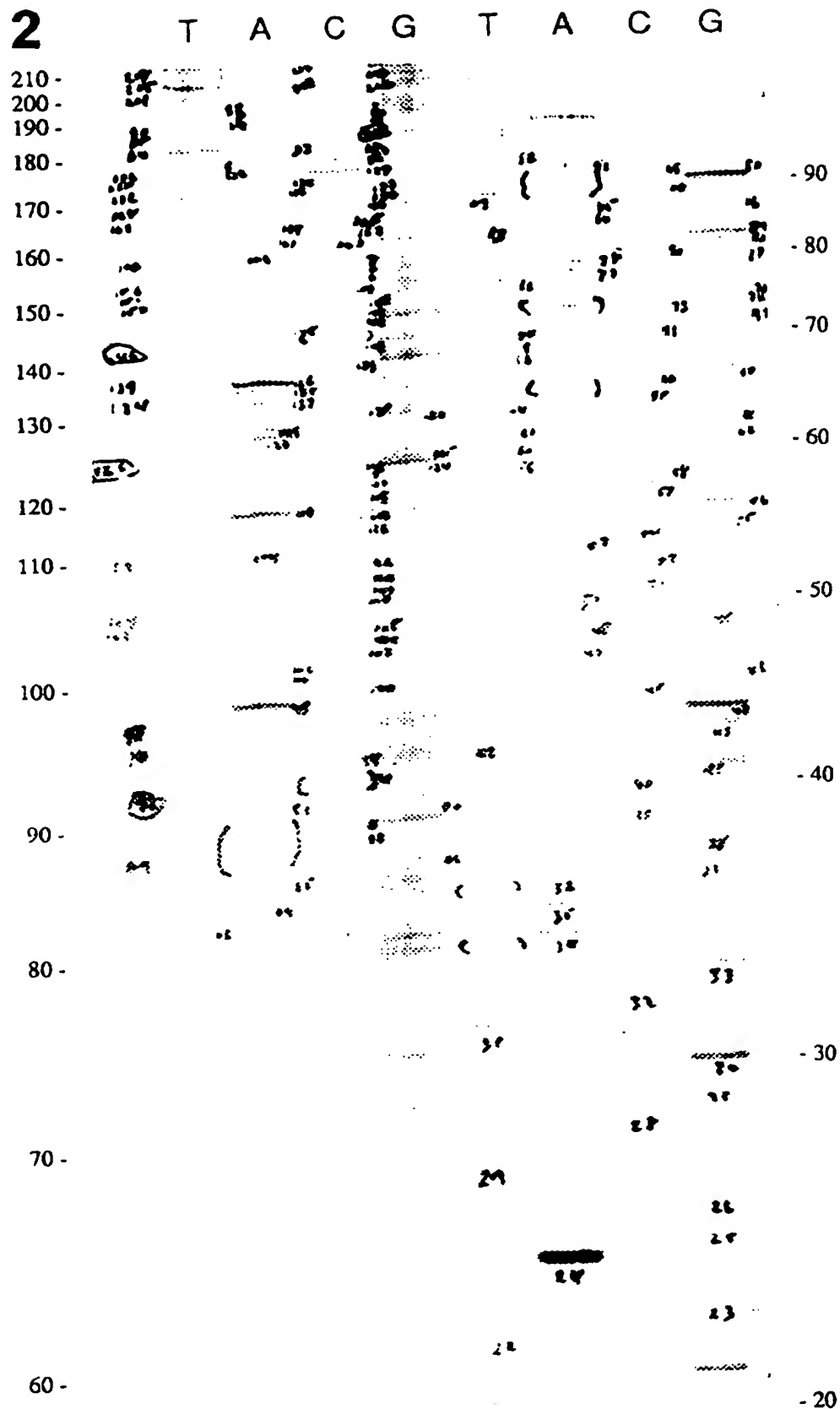
4. UNMODIFIED PRIMER 2 STRAND CAN BE DENATURED IN HOT EXO III BUFFER, THEN BIOTINYLATED PRIMER 1 STRAND CAN BE DETACHED FROM THE BEAD IN HOT LOADING BUFFER. BOTH FRAGMENTS CAN BE SEQUENCED SIMULTANEOUSLY BY PAGE.

PRIMER 1 STRAND				PRIMER 2 STRAND			
A	T	C	G	A	T	C	G
—		—	—		—	—	—
—		—	—	—	—	—	—
—	—	—	—	—	—	—	—
	—	—	—	—	—	—	—

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FIG. 2



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Figure 3 Legend. One-Step PCR Sequencing.

A portion of phage M13mp2 was sequenced by One-Step PCR Sequencing as described in the text. Primer SS20 was labeled and used, along with primer RP, for PCR amplification (250 bp product). Base-specific dNTPs were incorporated in four separate reactions and their positions were revealed by digestion with exonuclease II. The digested PCR products were separated by two loadings on a denaturing polyacrylamide gel.

As is shown in the first four lanes of Figure 2, the first loading separated fragments from position G₅₅ to about position A₂₁₀. The second loading separated fragments beginning at the first position beyond the 3' end of the primer, position G₂₁, to about position C₁₀₉.

On the figure, missing bands are denoted by circled numbers and extra bands are noted in parentheses.

The sequence for M13mp2 extending from position G₂₁ is:

GTGAGGTCGG	TCGAAAGGCC	GTGGCGAAGA	CCACGGCCTT
TGGTCCGTTT	CGCGGTAAGC	GGTAAGTCCG	ATGCGTTGAC
AACCCTTCCC	GCTAGCCACG	CCCGGAGAAG	CGATAATGCG
GTCGACCGCT	TTCCCCCTAC	ACGACGTTCC	GCTAATTCAA
CCCATTGCGG	TCCCAAAGG	GAAAAGGGTC	

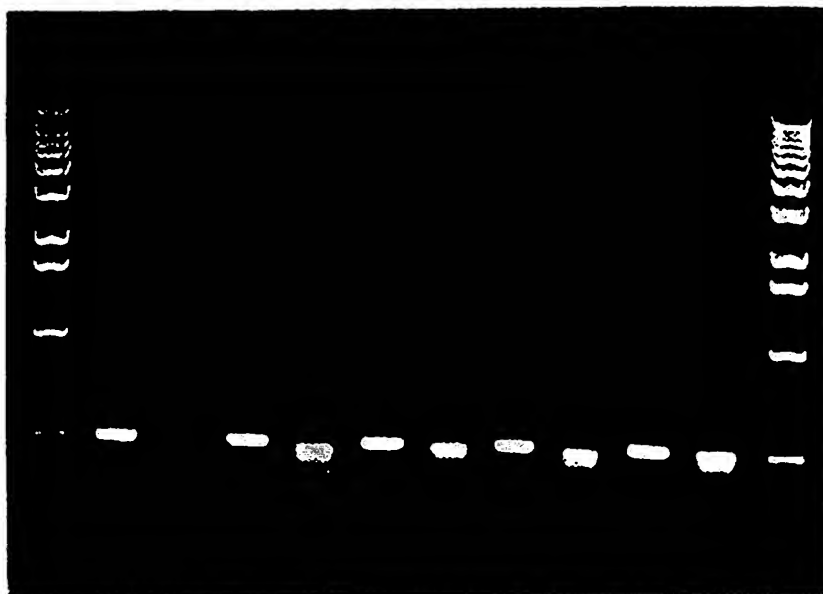
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dNTbp
Exo III

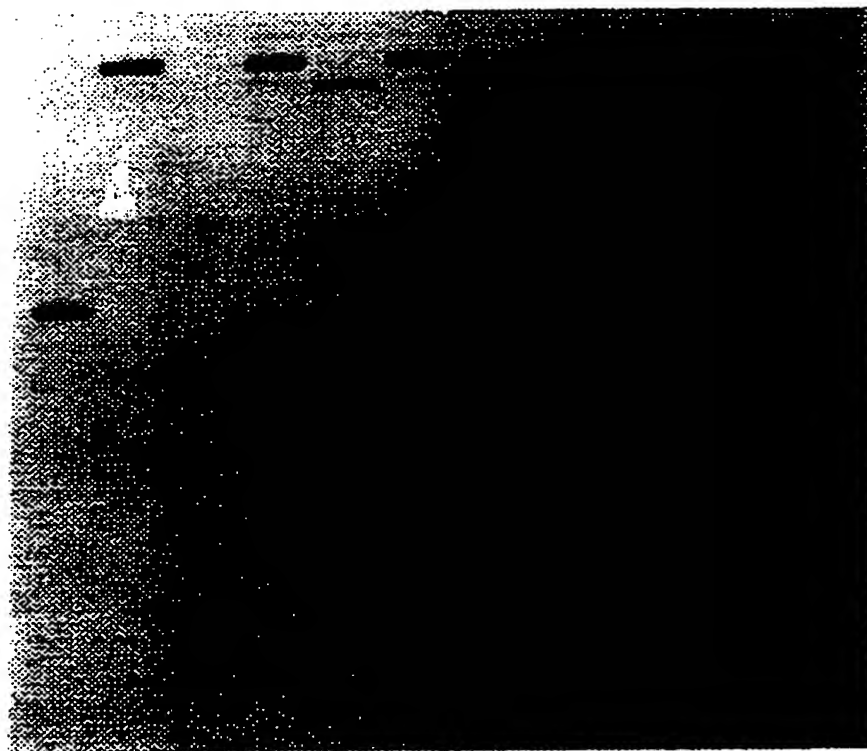
M	1	2	3	4	5	6	7	8	9	10	M
-	-	-	A	A	T	T	G	G	C	C	-
-	+	-	+	-	+	-	+	-	+	-	+

FIG. 4

dNTbp
Exo III

P	1	2	3	4	5	6	7	8	9	10	P
-	-	-	A	A	T	T	G	G	C	C	-
-	+	-	+	-	+	-	+	-	+	-	+

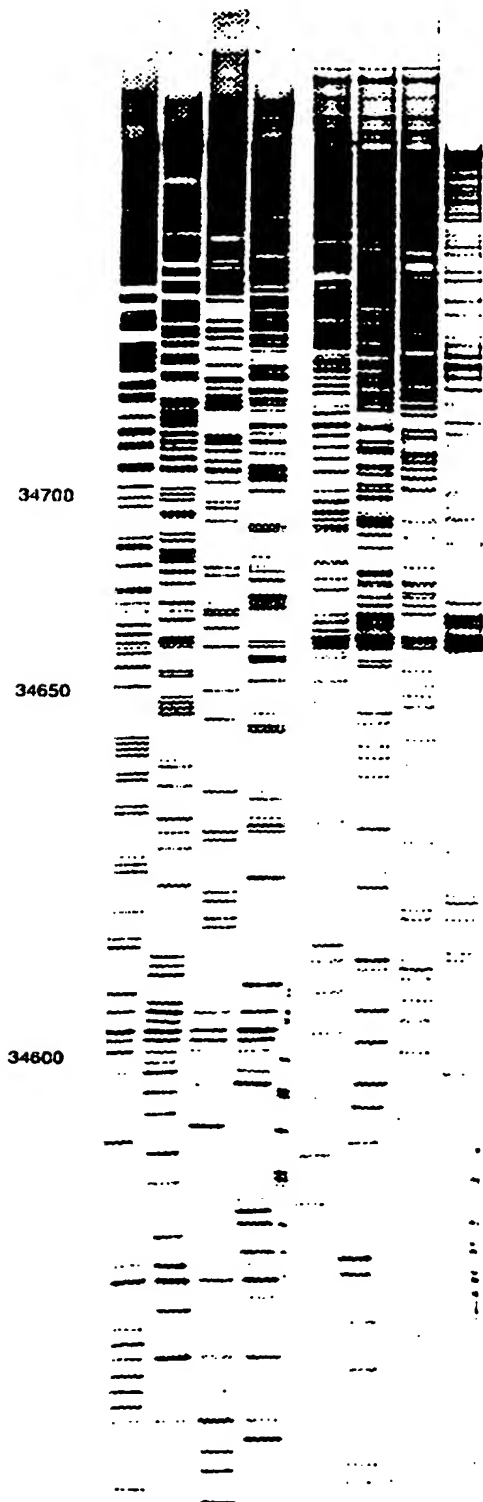
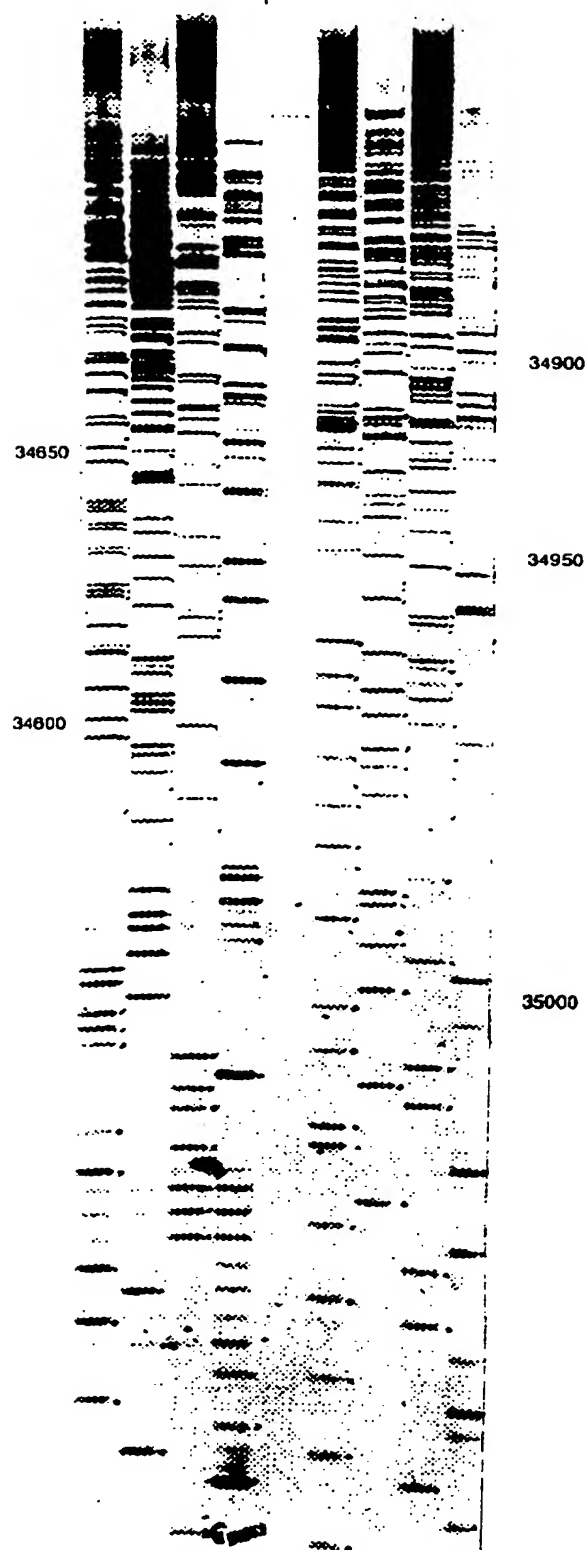
FIG. 5

Primer
Template

5' - CAGGAACAGCTATGGCCTC -3'
3' - GTCCTTGTCGATACCGGAGTCGATGTG -5'

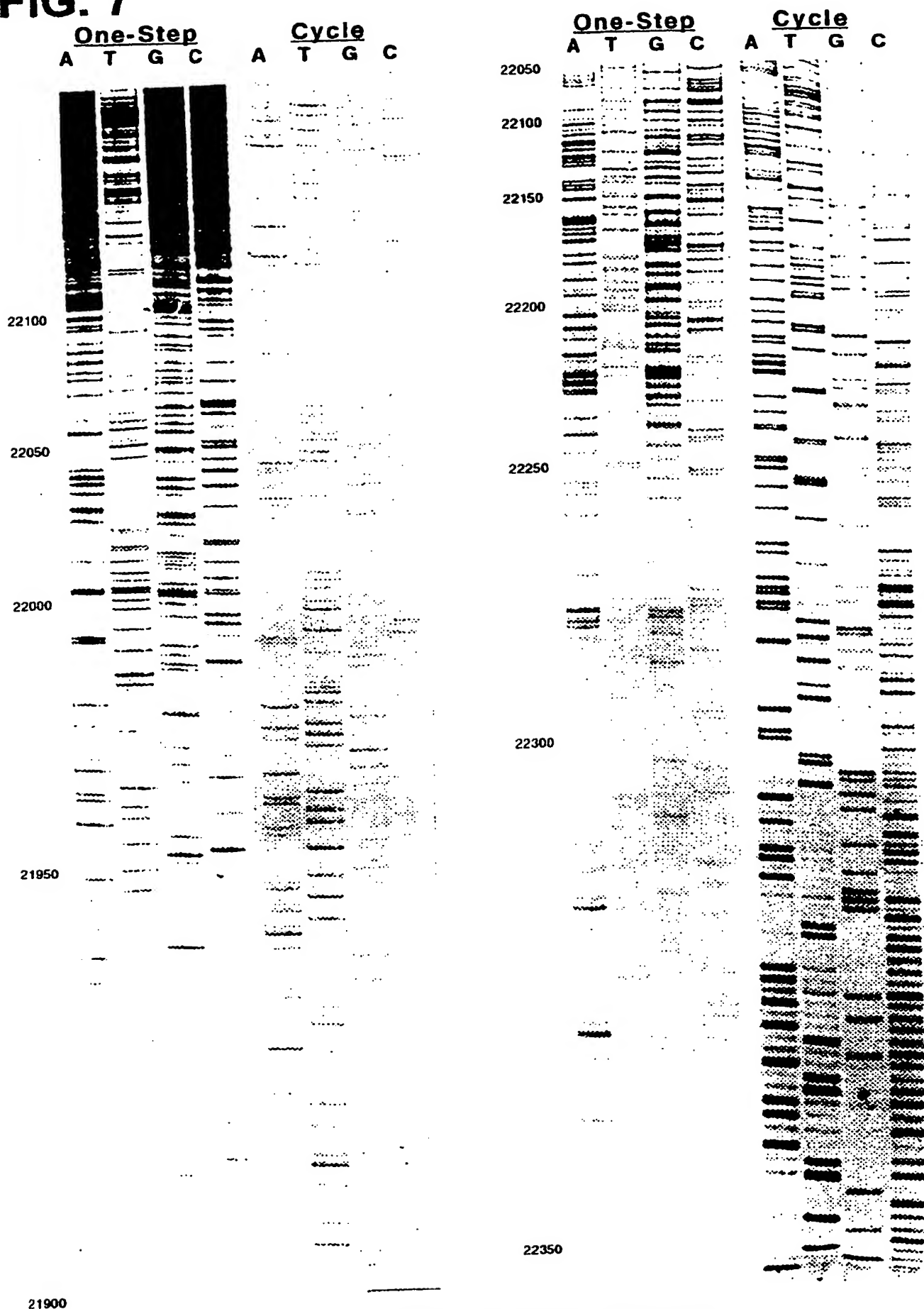
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FIG. 6Aforward reverse
T A C G T A C G**FIG. 6B**forward reverse
T A C G T A C G

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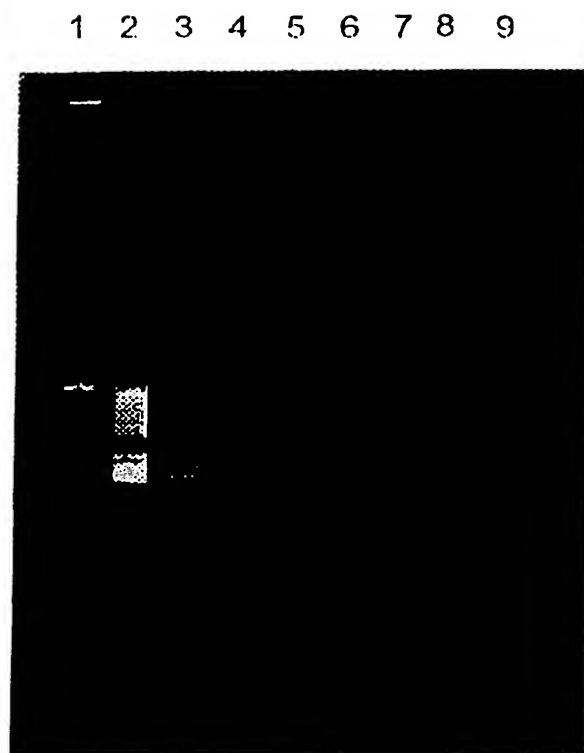
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FIG. 7

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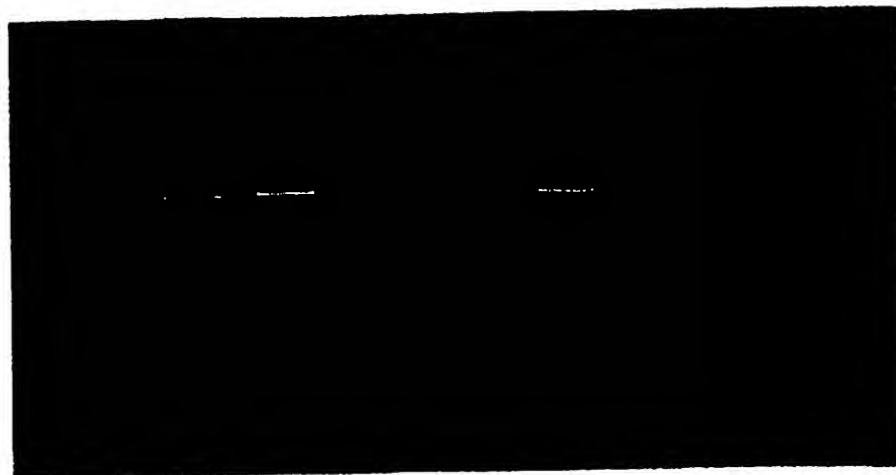
FIG. 8



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M 1 2 3 4 5 6

FIG. 9A



1 2 3 4 5 6 7 8 9

FIG. 9B



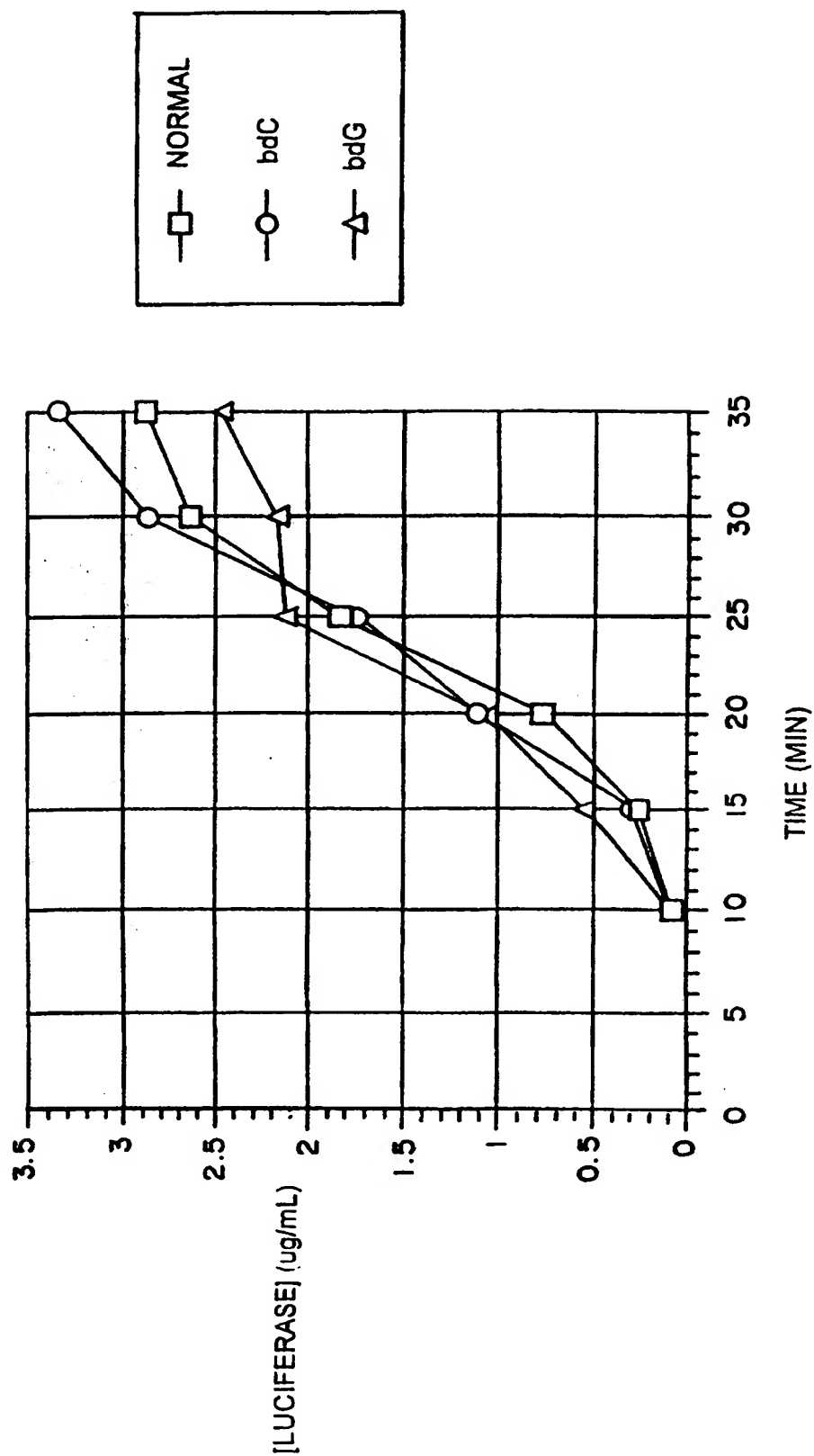
1 2 3

FIG. 9C



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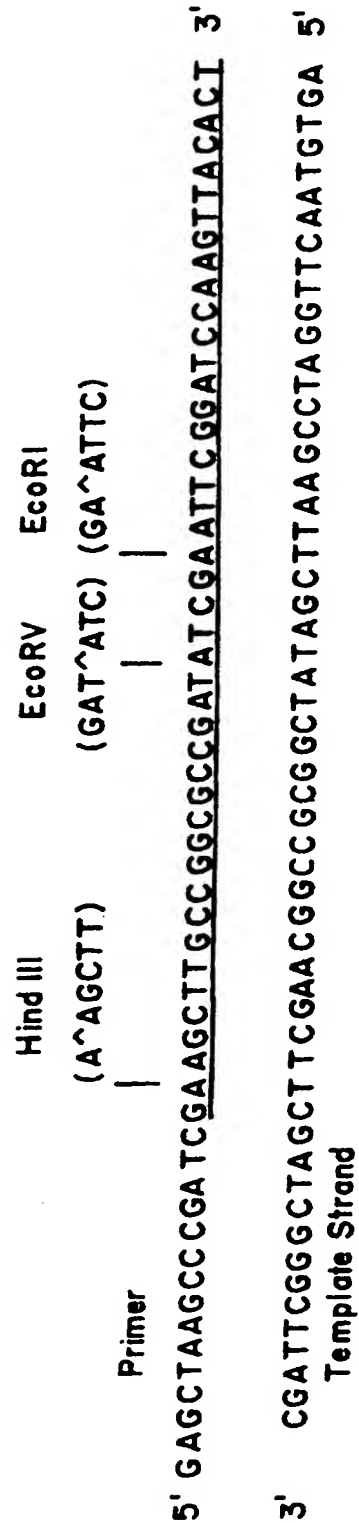
FIG. 10



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FIG. 13



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A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C07H 19/00, 21/00

US CL : 536/22.1, 4.1, 17.1, 18.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 536/22.1, 4.1, 17.1, 18.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG ONESEARCH: RIBONUCLEOSIDE BORANOPHOSPHATE, DEOXYRIBONUCLEOSIDE BORANOPHOSPHATE, OLIGORIBONUCLEOSIDE BORANOPHOSPHATE, OLIGODEOXYRIBONUCLEOTIDE BORANOPHOSPHATE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	5,177,198 A (SPIELVOGEL et al.) 05 January 1993, columns 7-8, Scheme 2; column 6, lines 40-41; column 2, lines 40-42	1, 5
Y		1-9
Y	US 4,672,110 A (LETSINGER) 09 June 1987, column 3, line 57-column 5, line 55	1-9

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

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P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

17 DECEMBER 1997

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28 JAN 1998

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